

"Peptides"

5 Summary of the invention

This invention relates to peptides which are fragments of protein products arising from frameshift mutations in genes, which peptides elicit T cellular immunity, and to cancer vaccines and compositions for anticancer treatment comprising said peptides.

10 The invention further relates to a method for identifying such peptides which are fragments of protein products arising from frameshift mutations in genes, which may elicit T cellular immunity which is useful for combating cancer associated with said mutated genes.

15 The invention also relates to DNA sequences encoding at least one frameshift mutant peptide, and vectors comprising at least one insertion site containing a DNA sequence encoding at least one frameshift mutant peptide.

20 Further the invention relates to methods for the treatment or prophylaxis of cancers associated with frameshift mutations in genes by administration of at least one frameshift mutant peptide or a recombinant virus vector comprising at least one insertion site containing a DNA sequence encoding at least one frameshift mutant peptide, or an isolated DNA sequence comprising a DNA sequence encoding at least one frameshift mutant peptide.

25 The present invention represents a further development of anticancer treatment or prophylaxis based on the use of peptides to generate activation and strengthening of the

anti cancer activity of the T cellular arm of the body's own immune system.

5 Technical Background

Tumour antigens, Status:

10 T cell defined antigens have now been characterised in a broad spectrum of cancer types. These antigens can be divided into several main groups, depending on their expression. The two main groups are constituted by developmental differentiation related antigens (tumour-testis antigens, oncofoetal antigens etc., such as MAGE antigens and CEA) and tissue specific differentiation antigens (Tyrosinase, gp100 etc.). The group containing the truly tumour specific antigens contains proteins that are altered due to mutations in the genes encoding them. In the majority of these, the mutations are unique and have been detected in a single or in a small number of tumours. Several of these antigens seem to play a role in oncogenesis.

20 Cancer vaccines, Status:

25 The focus in cancer vaccine development has been on antigens expressed in a high degree within one form of cancer (such as melanoma) or in many kinds of cancers. One reason for this is the increased recruitment of patients into clinical protocols. The field is in rapid growth, illustrated by the accompanying table listing the cancer vaccine protocols currently registered in the PDQ database of NCI.

Inheritable cancer/cancer gene testing:

5 Inherited forms of cancer occur at a certain frequency in the population. For several of these cancer forms, the underlying genetic defects have been mapped. This is also the case in Lynch syndrome cancers which constitute an important group of inheritable cancer. In families inflicted with this syndrome, family members inherit defect genes encoding DNA Mismatch Repair (MMR) Enzymes. Carriers of such MMR defects frequently develop colorectal cancer (HNPCC) and other forms of cancer (list?). Mutations in MMR enzymes can be detected using gene testing in the same way as other cancer related genes can be detected.

15 Gene testing of risk groups in this case represents an ethical dilemma, since no acceptable forms for prophylactic treatment exist. At present surgery to remove the organ in danger to develop cancer has been the only treatment option. In these patients, cancer vaccines will be a very (interesting) form of prophylaxis, provided efficient vaccines can be developed.

25 The lack of efficient repair of mismatched DNA results in deletions and insertions in one strand of DNA, and this happens preferentially in stretches of DNA containing repeated units (repeat sequences). Until now, focus has been on repeat sequences in the form of non-coding microsatellite loci. Indeed microsatellite instability is the hallmark of cancers resulting from MMR defects. We have taken another approach, and have concentrated on frameshift mutations occurring in DNA sequences coding for proteins related to the oncogenic process. Such frameshift mutations result in completely new amino acid sequences in the C-terminal part of the proteins, prematurely

terminating where a novel stop codon appears. This results in two important consequences:

- 1) The truncated protein resulting from the frameshift is generally nonfunctional, in most cases resulting in "knocking out" of an important cellular function. Aberrant proteins may also gain new functions such as the capacity to aggregate and form plaques. In both cases the frameshift results in disease.
- 2) The short new C-terminal amino acid sequence resulting from the shift in the reading frame (the "frameshift sequence") is foreign to the body. It does not exist prior to the mutation, and it only exists in cells having the mutation, i.e. in tumour cells and their pre malignant progenitors. Since they are completely novel and therefore foreign to the immune system of the carrier, they may be recognised by T-cells in the repertoire of the carrier. So far, nobody has focused on this aspect of frameshift mutations, and no reports exist on the characterisation of frameshift peptides from coding regions of proteins as tumour antigens. This concept is therefore novel and forms the basis for developing vaccines based on these sequences. It follows that such vaccines may also be used prophylactically in persons who inherit defective enzymes belonging to the MMR machinery. Such vaccines will therefore fill an empty space in the therapeutic armament against inherited forms of cancer.
- It has been shown that single amino acid substitutions in intracellular "self"-proteins may give rise to tumour rejection antigens, consisting of peptides differing in their amino acid sequence from the normal peptide. The T cells which recognise these peptides in the context of the major histocompatibility (MHC) molecules on the surface of the tumour cells, are capable of killing the

tumour cells and thus rejecting the tumour from the host.

In contrast to antibodies produced by the B cells, which typically recognise a free antigen in its native conformation and further potentially recognise almost any site exposed on the antigen surface, T cells recognise an antigen only if the antigen is bound and presented by a MHC molecule. Usually this binding will take place only after appropriate antigen processing, which comprises a proteolytic fragmentation of the protein, so that the resulting peptide fragment fits into the groove of the MHC molecule. Thereby T cells are enabled to also recognise peptides derived from intracellular proteins. T cells can thus recognise aberrant peptides derived from anywhere in the tumour cell, in the context of MHC molecules on the surface of the tumour cell, and can subsequently be activated to eliminate the tumour cell harbouring the aberrant peptide.

- 20 M. Barinaga, Science, 257, 880-881, 1992 offers a short review of how MHC binds peptides. A more comprehensive explanation of the Technical Background for this Invention may be found in D. Male et al, Advanced Immunology, 1987, J.B. Lippincott Company, Philadelphia.
- 25 Both references are hereby included in their entirety.

The MHC molecules in humans are normally referred to as HLA (human leukocyte antigen) molecules. They are encoded by the HLA region on the human chromosome No 6.

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The HLA molecules appear as two distinct classes depending on which region of the chromosome they are encoded by and which T cell subpopulations they interact with and thereby activate primarily. The class I molecules are encoded by the HLA A, B and C subloci and they primarily activate CD8+ cytotoxic T cells. The HLA

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class II molecules are encoded by the DR, DP and DQ subloci and primarily activate CD4+ T cells, both helper cells and cytotoxic cells.

- 5 Normally every individual has six HLA Class I molecules, usually two from each of the three groups A, B and C. Correspondingly, all individuals have their own selection of HLA Class II molecules, again two from each of the three groups DP, DQ and DR. Each of the groups A, B, C
10 and DP, DQ and DR are again divided into several subgroups. In some cases the number of different HLA Class I or II molecules is reduced due to the overlap of two HLA subgroups.
- 15 All the gene products are highly polymorphic. Different individuals thus express distinct HLA molecules that differ from those of other individuals. This is the basis for the difficulties in finding HLA matched organ donors in transplantations. The significance of the genetic
20 variation of the HLA molecules in immunobiology is reflected by their role as immune-response genes. Through their peptide binding capacity, the presence or absence of certain HLA molecules governs the capacity of an individual to respond to peptide epitopes. As a
25 consequence, HLA molecules determine resistance or susceptibility to disease.

T cells may control the development and growth of cancer by a variety of mechanisms. Cytotoxic T cells, both HLA
30 class I restricted CD8+ and HLA Class II restricted CD4+, may directly kill tumour cells carrying the appropriate tumour antigens. CD4+ helper T cells are needed for cytotoxic CD8+ T cell responses as well as for antibody responses, and for inducing macrophage and LAK cell
35 killing.

A requirement for both HLA class I and II binding is that the peptides must contain a binding motif, which usually is different for different HLA groups and subgroups. A binding motif is characterised by the requirement for amino acids of a certain type, for instance the ones carrying large and hydrophobic or positively charged side groups, in definite positions of the peptide so that a narrow fit with the pockets of the HLA binding groove is achieved. The result of this, taken together with the peptide length restriction of 8-10 amino acids within the binding groove, is that it is quite unlikely that a peptide binding to one type of HLA class I molecules will also bind to another type. Thus, for example, it may very well be that the peptide binding motif for the HLA-A1 and HLA-A2 subgroups, which both belong to the class I group, are as different as the motifs for the HLA-A1 and HLA-B1 molecules.

For the same reasons it is not likely that exactly the same sequence of amino acids will be located in the binding groove of the different class II molecules. In the case of HLA class II molecules the binding sequences of peptides may be longer, and it has been found that they usually contain from 10 to 16 amino acids, some of which, at one or both terminals, are not a part of the binding motif for the HLA groove.

However, an overlap of the different peptide binding motifs of several HLA class I and class II molecules may occur. Peptides that have an overlap in the binding sequences for at least two different HLA molecules are said to contain "nested T cell epitopes". The various epitopes contained in a "nested epitope peptide" may be formed by processing of the peptide by antigen presenting cells and thereafter be presented to T cells bound to different HLA molecules. The individual variety of HLA

molecules in humans makes peptides containing nested epitopes more useful as general vaccines than peptides that are only capable of binding to one type of HLA molecule.

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Effective vaccination of an individual can only be achieved if at least one type of HLA class I and/or II molecule in the patient can bind a vaccine peptide either in it's full length or as processed and trimmed by the patient's own antigen presenting cells.

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The usefulness of a peptide as a general vaccine for the majority of the population increases with the number of different HLA molecules it can bind to, either in its full length or after processing by antigen presenting cells.

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In order to use peptides derived from a protein encoded by a mutated gene as vaccines or anticancer agents to generate anti tumour CD4+ and/or CD8+ T cells, it is necessary to investigate the mutant protein in question and identify peptides that are capable, eventually after processing to shorter peptides by the antigen presenting cells, to stimulate T cells.

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Prior art

In our International Application PCT/NO92/00032 (published as WO92/14756), we described synthetic peptides and fragments of oncogene protein products which have a point of mutation or translocations as compared to their proto-oncogene or tumour suppressor gene protein. These peptides correspond to, completely cover or are fragments of the processed oncogene protein fragment or tumour suppressor gene fragment as presented by cancer

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cells or other antigen presenting cells, and are presented as a HLA-peptide complex by at least one allele in every individual. These peptides were also shown to induce specific T cell responses to the actual oncogene protein fragment produced by the cell by processing and presented in the HLA molecule. In particular, we described peptides derived from the p21 ras protein which had point mutations at particular amino acid positions, namely position 12, 13 and 61. These peptides have been shown to be effective in regulating the growth of cancer cells *in vitro*. Furthermore, the peptides were shown to elicit CD4+ T cell immunity against cancer cells harbouring the mutated p21 ras oncogene protein through the administration of such peptides in vaccination or cancer therapy schemes. Later we have shown that these peptides also elicit CD8+ T cell immunity against cancer cells harbouring the mutated p21 ras oncogene protein through the administration mentioned above.

However, the peptides described above will be useful only in certain number of cancers, namely those which involve oncogenes with point mutations or translocation in a proto-oncogene or tumour suppressor gene. There is therefore a strong need for an anticancer treatment or vaccine which will be effective against a more general range of cancers.

In general, tumors are very heterogenous with respect to genetic alterations found in the tumour cells. This implies that both the potential therapeutic effect and prophylactic strength of a cancer vaccine will increase with the number of targets that the vaccine is able to elicit T cell immunity against. A multiple target vaccine will also reduce the risk of new tumour formation by treatment escape variants from the primary tumour.

Definition of Problem solved by the Invention.

There is a continuing need for new anticancer agents based on antigenic peptides giving rise to specific T cellular responses and toxicity against tumours and cancer cells carrying genes with mutations related to cancer. The present invention will contribute largely to supply new peptides that can have a use in the combat and prevention of cancer as ingredients in a multiple target anti-cancer vaccine.

Another problem solved by the present invention is that a protection or treatment can be offered to the individuals belonging to family's or groups with high risk for hereditary cancers. Hereditary cancers are in many cases associated with genes susceptible to frameshift mutations as described in this invention (i.e. mutations in mismatch repair genes). Today it is possible to diagnose risk of getting hereditary cancer but no pharmaceutical method for protection against the onset of the cancer is available.

Definition of the Invention

A main object of the invention is to obtain peptides corresponding to peptide fragments of mutant proteins produced by cancer cells which can be used to stimulate T cells.

Another main object of the invention is to develop a cancer therapy for cancers based on the T cell immunity which may be induced in patients by stimulating their T cells either *in vivo* or *in vitro* with the peptides according to the invention.

A third main object of the invention is to develop a vaccine to prevent the establishment of or to eradicate cancers based solely or partly on peptides corresponding to peptides of the present invention which can be used to generate and activate T cells which produce cytotoxic T cell immunity against cells harbouring the mutated genes.

A fourth main object of the invention is to design an anticancer treatment or prophylaxis specifically adapted to a human individual in need of such treatment or prophylaxis, which comprises administering at least one peptide according to this invention.

These and other objects of the invention are achieved by the attached claims.

Since frameshift mutations result in premature stop codons and therefore deletion in large parts of the proteins, proteins with frameshift mutations have generally not been considered to be immunogenetic and have therefore not been considered as targets for immunotherapy. Thus it has now surprisingly been found that a whole group of new peptides resulting from frameshift mutations in tumour relevant genes are useful for eliciting T cell responses against cancer cells harbouring genes with such frameshift mutations.

Genes containing a mono nucleoside base repeat sequence of at least five residues, for example of eighth deoxyadenosine bases (AAAAAAAA), or a di-nucleoside base repeat sequence of at least four di-nucleoside base units, for example of two deoxyadenosine-deoxycytosine units (ACAC), are susceptible to frameshift mutations. The frameshift mutations occur, respectively, either by insertion of one or two of the mono-nucleoside base residue or of one or two of the di-nucleoside base unit

in the repeat sequence, or by deletion of one or two of the mono-nucleoside base residue or of one or two of the di-nucleoside base unit from the repeat sequence. A gene with a frameshift mutation will from the point of mutation code for a protein with a new and totally different amino acid sequence as compared to the normal gene product. This mutant protein with the new amino acid sequence at the carboxy end will be specific for all cells carrying the modified gene.

In the remainder of this specification and claims the denomination frameshift mutant peptides will comprise such proteins and peptide fragments thereof.

It has now according to the present invention been found that such new protein sequences arising from frameshift mutations in genes in cancer cells give rise to tumour rejection antigens that are recognised by T cells in the context of HLA molecules.

It has further according to the present invention been found a group of peptides corresponding to fragments of mutant proteins arising from frameshift mutations in genes in cancer cells which can be used to generate T cells. The said peptides can therefore also be used to rise a T cell activation against cancer cells harbouring a gene with a frameshift mutation as described above.

These peptides are at least 8 amino acids long and correspond, either in their full length or after processing by antigen presenting cells, to the mutant gene products or fragments thereof produced by cancer cells in a human patient afflicted with cancer.

A peptide according to this invention is characterised in that it

a) is at least 8 amino acids long and is a fragment of a mutant protein arising from a frameshift mutation in a gene of a cancer cell;

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and

b) consists of at least one amino acid of the mutant part of a protein sequence encoded by said gene;

10

and

c) comprises 0-10 amino acids from the carboxyl terminus of the normal part of the protein sequence preceding the amino terminus of the mutant sequence and may further extend to the carboxyl terminus of the mutant part of the protein as determined by a new stop codon generated by the frameshift mutation in the gene;

15

20

and

d) induces, either in its full length or after processing by antigen presenting cell, T cell responses.

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The peptides of this invention contain preferably 8-25, 9-20, 9-16, 8-12 or 20-25 amino acids. They may for instance contain 9, 12, 13, 16 or 21 amino acids.

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It is most preferred that the peptides of the present invention are at least 9 amino acids long, for instance 9-18 amino acids long, but due to the processing

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possibility of the antigen presenting cells also longer peptides are very suitable for the present invention. Thus the whole mutant amino acid sequence may be used as a frameshift mutant peptide according to the present invention, if it comprises 8 amino acids or more.

The invention further relates to a method for vaccination of a person disposed for cancer, associated with a frameshift mutation in a gene, consisting of administering at least one peptide of the invention one or more times in an amount sufficient for induction of T-cell immunity to the mutant proteins encoded by the frameshift mutated gene.

The invention also relates to a method for treatment of a patient afflicted with cancer associated with frameshift mutation in genes, consisting of administering at least one peptide of the invention one or more times in an amount sufficient for induction of T-cell immunity to mutant proteins arising from frameshift mutations in the genes of cancer cells.

Furthermore, it has according to the present invention been found a method for identifying new peptides which correspond to fragments of proteins arising from frameshift mutations in genes. This method is characterised by the following steps:

- 1) identifying a gene in a cancer cell susceptible to frameshift mutation by having a mono nucleoside base repeat sequence of at least five residues, or a di-nucleoside base repeat sequence of at least four di-nucleoside base units;

and

2) removing, respectively, one nucleoside base
residue or one di-nucleoside base unit from the
repeat sequence and identifying the amino acid
sequence of the protein encoded by the altered gene
sequence as far as to include a new stop codon;

5

and/or

3) removing, respectively, two nucleoside base
residues or two di-nucleoside base units from the
repeat sequence and identifying the amino acid
sequence of the protein encoded by the altered gene
sequence as far as to include a new stop codon;

10

and/or

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4) inserting, respectively, one nucleoside base
residue or one di-nucleoside base unit in the
repeat sequence and identifying the amino acid
sequence of the protein encoded by the altered gene
sequence as far as to include a new stop codon;

20

and/or

5) inserting, respectively, two nucleoside base
residues or two di-nucleoside base units in the
repeat sequence and identifying the amino acid
sequence of the protein encoded by the altered gene
sequence as far as to include a new stop codon.

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In order to determine whether the peptides thus
identified are useable in the compositions and methods
according to the present invention for the treatment or
prophylaxis of cancer, the following further step should
be performed:

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6) determining whether the new peptide, either in their full length or as shorter fragments of the peptides, are able to stimulate T cells.

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Optionally a further step may be added as follows:

7) determining peptides containing nested epitopes for different major HLA class I and/or HLA class II molecules.

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Detailed Description of the invention.

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In the present description and claims, the amino acids are represented by their one letter abbreviation as known in the art.

20 The peptides of the present invention shall be explicitly exemplified through two different embodiments, wherein cancer develops based on frameshift mutations in specific genes, namely the BAX gene and TGF β RII gene:

25 I) BAX gene

It has been established that the BAX gene is involved in regulation of survival or death of cells by promoting apoptosis. The human BAX gene contains a repeat sequence of eight deoxyguanosine bases (G8) in the third exon, spanning codons 38 to 41 (ATG GGG GGG GAG).

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Frameshift mutations in this G8 repeat have been observed, both as G7 (ATG GGG GGG AGG) and G9 (ATG GGG GGG GGA) repeats, both in colon cancer cells and prostate cancer cells. The occurrence is more than 50% of the examined cases (Rampino, N. et al., "Somatic frameshift

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mutations in the BAX gene in colon cancers of the
 microsatellite mutator phenotype.", Science (Washington
 DC), 275: 967-969, 1997). The modified BAX gene products
 are unable to promote apoptosis and thus makes further
 5 tumour progress possible. Furthermore the modified gene
 products are only found in cancer cells and are therefore
 targets for specific immunotherapy.

According to the present invention, peptides
 10 corresponding to the transformed BAX protein products
 arising from frameshift mutations in the BAX gene can be
 used as anticancer therapeutical agents or vaccines with
 the function to trigger the cellular arm of the immune
 system (T-cells) against cancer cells in patients
 15 afflicted with cancers associated with a modified BAX
 gene.

Frameshift mutations in the BAX gene result in mutant
 peptide sequences with the first amino acid of the
 20 altered sequence in position 41 as compared to the normal
 BAX protein (Table 1, seq.id. no. 1 to 4).

Table 1

amino acid pos	41	51	61	71
25 normal bax peptide ;	EAP	EL	ALDPV	PQDASTKKLS ECLKRIGDEL
DS...				
seq.id.no. 1(bax-1G);	RHPSWPWTRC	LRMRPPRS		
seq.id.no. 4(bax+2G);	GRHPSWPWTR	CLMRPPRS		
seq.id.no. 2(bax-2G);	GTRAGPGPGA	SGCVHQEAER	VSQAHRGRTG	Q
30 seq.id.no. 3(bax+1G);	GGTRAGPGPG	ASGCVHQEAE	RVSQAHRGRT	GQ

Table 2 shows one group of peptides according to the present invention:

Table 2

- 5 seq.id.no. 5: IQDRAGRMGGRHPSWPWTRCLMRPPRS
 seq.id.no. 6: IQDRAGRMGGGRHPSWPWT
 seq.id.no. 7: IQDRAGRMGGGGTRAGPGPGASGCVHQEAERVSQAHRGRTGQ
 seq.id.no. 8: IQDRAGRMGGGTRAGPGPG

- 10 The peptides listed in Table 3 were used for *in vitro* generation of T cells that recognise mutant BAX peptides.

Table 3.

- seq id no 1: RHPSWPWTRCLMRPPRS
 15 seq id no 9: IQDRAGRMGGRHPSWPWTRCLR
 seq id no 6: IQDRAGRMGGGRHPSWPWT
 seq id no 10: ASGCVHQEAERVSQAHRGRTGQ
 seq id no 11: GGTRAGPGPGASGCVHQEAERV
 seq id no 12: IQDRAGRMGGGGTRAGPGPGAS
 20 seq id no 8: IQDRAGRMGGGTRAGPGPG

The most preferred peptides according to this embodiment of the present invention are listed in Table 4:

25 Table 4

- seq id no 1: RHPSWPWTRCLMRPPRS
 seq id no 2: GTRAGPGPGASGCVHQEAERVSQAHRGRTGQ
 seq id no 3: GGTRAGPGPGASGCVHQEAERVSQAHRGRTGQ
 seq id no 4: GRHPSWPWTRCLMRPPRS
 30 seq.id.no. 5: IQDRAGRMGGRHPSWPWTRCLMRPPRS
 seq.id.no. 6: IQDRAGRMGGGRHPSWPWT
 seq.id.no. 7: IQDRAGRMGGGGTRAGPGPGASGCVHQEAERVSQAHRGRTGQ
 seq id no 8: IQDRAGRMGGGTRAGPGPG
 seq id no 9: IQDRAGRMGGRHPSWPWTRCLR
 35 seq id no 10: ASGCVHQEAERVSQAHRGRTGQ
 seq id no 11: GGTRAGPGPGASGCVHQEAERV

seq id no 12: IQDRAGRMGGGGTRAGPGPGAS

2) TGF β RII

It has been established that the TGF β RII gene is involved
5 in regulation of cell growth. TGF β RII is a receptor for
TGF β which down regulates cell growth. The human gene
coding for TGF β RII contains a repeat sequence of ten
deoxyadenosine bases (A10) from base no. 709 to base no.
718 (GAA AAA AAA AAG CCT). In colon cancers and
10 pancreatic cancers frameshift mutations in this A10
repeat have been observed, both as A9 (GAA AAA AAA AGC
CT) and A11 (GAA AAA AAA AAA GCC) repeats, in
approximately 80 % of the examined cases (Yamamoto, H.,
"Somatic frameshift mutations in DNA mismatch repair and
15 proapoptosis genes in hereditary nonpolyposis colorectal
cancer.", Cancer Research 58, 997-1003, March 1, 1998).
The modified TGF β RII gene products are unable to bind
TGF β and the signal for down regulation of cell growth
is eliminated and thus makes further tumour progress
20 possible. Furthermore the modified gene products are only
found in cancer cells and are therefore targets for
immunotherapy.

Consequently peptides corresponding to the transformed
25 TGF β RII protein products arising from frameshift
mutations in the TGF β RII gene can be used as anticancer
therapeutical agents or vaccines with the function to
trigger the cellular arm of the immune system (T-cells)
against cancer cells in patients afflicted with cancers
30 associated with a modified TGF β RII gene.

Frameshift mutations in the TGF β RII gene result in mutant
peptide sequences with the first amino acid of the
altered sequence in either position 133 (one and two base

deletions) or 134 (one and two base insertions) as compared to the normal TGF β RII protein (Table 5, seq.id.nos. 13 and 21).

5 Table 5.

amino acid pos. 133

normal TGF β RII ; K PGETFFMCSC SSDECNDNII FSEEYNTSNP
DLLL

seq id no 13(-1A); S LVRLSSCVPV ALMSAMTTSS SQKNITPAIL TCC

10 seq id no 13(+2A); SLVRLSSCVP VALMSAMTTS SSQKNITPAI
LTCC

TGF β RII + 1A) ; AW

TGF β RII - 2A) ; A W

15 Table 6 shows one groups of peptides of this invention:

Table 6

seq id no 14:SPKCIMKEKKSLVRLSSCVPVALMSAMTTSSSQKNITPAILTCC

seq id no 15:PKCIMKEKKKSLVRLSSCV

20 seq id no 19:SPKCIMKEKKAW

seq id no 20:PKCIMKEKKKAW

Table 7 presents peptides that were used for *in vitro* generation of T cells that recognise mutant TGF β RII peptides.

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Table 7

seq id no 15: PKCIMKEKKKSLVRLSSCV

seq id no 16: ALMSAMTTSSSQKNITPAILTCC

30 seq id no 17: SLVRLSSCVPVALMSAMTTSSSQ

seq id no 18: SPKCIMKEKKSLVRLSSCVPVA

seq id no 19: SPKCIMKEKKAW

seq id no 20: PKCIMKEKKKAW

seq id no 21: AMTTSSSQKNITPAILTCC

35 seq id no 428: SLVRLSSCV

The most preferred peptides of this embodiment of the present invention are:

5 Table 8

- seq id no 13:SLVRLSSCVPVALMSAMTTSSSQKNITPAILTCC
seq id no 14:SPKCIMKEKKSLVRLSSCVPVALMSAMTTSSSQKNITPAILTCC
seq id no 15:PKCIMKEKKKSLVRLSSCV
seq id no 16:ALMSAMTTSSSQKNITPAILTCC
10 seq id no 17:SLVRLSSCVPVALMSAMTTSSSQ
seq id no 18:SPKCIMKEKKSLVRLSSCVPVA
seq id no 19:SPKCIMKEKKAW
seq id no 20:PKCIMKEKKAW
seq id no 21:AMTTSSSQKNITPAILTCC
15 seq id no428:SLVRLSSCV

Other peptides of the invention can be fragments of the peptides listed in the Tables 1-8 above. Such fragments are most preferred from 9-16 amino acids long and include
20 at least one amino acid from the mutant part of the protein.

As used in this description and claims the term fragment is intended to specify a shorter part of a longer peptide
25 or of a protein.

Other cancer associated genes containing repeat sequences of a nucleoside base and which therefore are susceptible to frameshift mutations and consequently are potential
30 candidates for peptides according to the present invention (seq id nos according to table 9 are given in parentheses in each case) are the following:

- Human TGF- β -2 (hTGF β 2) gene (seq id nos 22-29)
35 Deleted in colorectal cancer (DCC) gene (seq.id.nos. 30-34)

- Human breast and ovarian cancer susceptibility (BRCA1) gene (seq.id.nos. 378-387)
- Human breast cancer susceptibility (BRCA2) gene (seq.id.nos. 35-94)
- 5 Human protein tyrosine phosphatase (hPTP) gene (seq.id.nos. 95-102)
- Human DNA topoisomerase II (top2) gene (seq.id.nos. 103-108)
- Human kinase (TTK) gene (seq.id.nos. 109-120)
- 10 Human transcriptional repressor (CTCF) gene (seq.id.nos. 121-127)
- Human FADD-homologous ICE/CED-3-like protease gene (seq.id.nos. 128-133)
- Human putative mismatch repair/binding protein (hMSH3) gene (seq.id.nos. 134-147)
- 15 Human retinoblastoma binding protein 1 isoform I (hRBP1) gene (seq.id.nos. 148-156)
- Human FMR1 (hFMR1) gene (seq.id.nos. 157-161)
- Human TINUR gene (seq.id.nos. 162-169)
- 20 b-raf oncogene (seq.id.nos. 170-175)
- Human neurofibromin (NF1) gene (seq.id.nos. 176-181)
- Human germline n-myc gene (seq.id.nos. 182-188)
- Human n-myc gene (seq.id.nos. 189-194)
- Human ras inhibitor gene (seq.id.nos. 195-199)
- 25 Human hMSH6 gene (seq.id.nos. 200-203 and 293-297)
- Human nasopharynx carcinoma EBV BNLF-1 gene (seq.id.nos. 204-210)
- Human cell cycle regulatory protein (E1A-binding protein) p300 gene (seq.id.nos. 211-218)
- 30 Human B-cell lymphoma 3-encoded protein (bcl-3) gene (seq.id.nos. 219-226)
- Human transforming growth factor-beta induced gene product (BIGH3) (seq.id.nos. 227-232)
- Human transcription factor ETV1 gene (seq.id.nos. 233-239)
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- Human insulin-like growth factor binding protein (IGFBP4) gene (seq.id.nos. 240-246)
- Human MUC1 gene (seq.id.nos. 247-266)
- Human protein-tyrosine kinase (JAK1) gene (seq.id.nos. 267-271)
- Human protein-tyrosine kinase (JAK3) gene (seq.id.nos. 272-279)
- Human Flt4 gene (for transmembrane tyrosinase kinase) (seq.id.nos. 280-284)
- Human p53 associated gene (seq.id.nos. 285-292)
- Human can (hCAN) gene (seq.id.nos. 298-300)
- Human DBL (hDBL) proto-oncogene / Human MCF2PO (hMCF2PO) gene (seq.id.nos. 301-306)
- Human dek (hDEK) gene (seq.id.nos. 307-309)
- Human retinoblastoma related protein (p107) gene (seq.id.nos. 310-313)
- Human G protein-coupled receptor (hGPR1) gene (seq.id.nos. 314-319)
- Human putative RNA binding protein (hRBP56) gene (seq.id.nos. 320-325)
- Human transcription factor (hITF-2) gene (seq.id.nos. 326-327)
- Human malignant melanoma metastasis-supressor (hKiSS-1) gene (seq.id.nos. 328-334)
- Human telomerase-associated protein TP-1 (hTP-1) gene (seq.id.nos. 335-348)
- Human FDF-5 (hFDF-5) gene (seq.id.nos. 349-356)
- Human metastasis-assosiated mtal (hMTA1) gene (seq.id.nos. 357-362)
- Human transcription factor TFIIB 90 kDa subunit (hTFIIB90) gene (seq id nos 363-369)
- Human tumour suppressor (hLUCA-1) gene (seq id nos 370-377)
- Human Wilm's tumour (WIT-1) associated protein (seq id nos 388-393)

Human cysteine protease (ICERel-III) gene (seq id nos 394-398 and 459)

Human Fas ligand (FasL) gene (seq id nos 399-403)

Human BRCA1-associated RING domain protein (BARD1) gene
5 (seq id nos 404-417)

Human mcf.2 (hMCF.2) gene (seq id nos 418-422)

Human Fas antigen (fas) gene (seq id nos 423-427)

Human DPC4 gene (seq id nos 429-437).

10 The mutant peptides that are the results of frameshift mutation in these genes, in accordance with the present invention, are listed in table 9.

Table 9

15	seq id no 22; TVGRPHISC
	seq id no 23; KTVGRPHISC
	seq id no 24; KQWEDPTSPANVIALLOT
	seq id no 25; QWEDPTSPANVIALLOT
	seq id no 26; QKTIKSTRKKTVGRPHISC
20	seq id no 27; QKTIKSTRKKKTVGRPHISC
	seq id no 28; QKTIKSTRKKKQWEDPTSPANVIALLOT
	seq id no 29; QKTIKSTRKKQWEDPTSPANVIALLOT
	seq id no 30; AADLQQQFVHFLDCWDVSSIPFTLHLPQAQDITT
	seq id no 31; GKDAKEKSS
25	seq id no 32; GKDAKEKKSS
	seq id no 33; GKDAKEKKAADLQQQFVHFLDCWDVSSIPFTLHLPQAQDITT
	seq id no 34; GKDAKEKAADLQQQFVHFLDCWDVSSIPFTLHLPQAQDITT
	seq id no 35; FSMKQTLNMVKNLGTK
	seq id no 36; KFSMKQTLNMVKNLGTK
30	seq id no 37; VRTSKTRKKFSMKQTLNMVKNLGTK
	seq id no 38; VRTSKTRKKKFSMKQTLNMVKNLGTK
	seq id no 39; VRTSKTRKKNFP
	seq id no 40; VRTSKTRKKNFP
	seq id no 41; IKKKLLQFQK
35	seq id no 42; KIKKKLLQFQK
	seq id no 43; KSRRNYFNFKNQCQSR

seq id no 44; SRRNYFNFKNNCQSRL
seq id no 45; TNLRVIQKIKKKLLQFQK
seq id no 46; TNLRVIQKKIKKKLLQFQK
seq id no 47; TNLRVIQKKSRRNYFNFKNNCQSRL
5 seq id no 48; TNLRVIQKSRRNYFNFKNNCQSRL
seq id no 49; KIMIT
seq id no 50; NIDKIKEKIMIT
seq id no 51; NIDKIKEKKIMIT
seq id no 52; IINAN
10 seq id no 53; KIINAN
seq id no 54; NDKTVSEKIINAN
seq id no 55; NDKTVSEKKIINAN
seq id no 56; NGLEKEYLMVNQKE
seq id no 57; SQTSLLEAKNGLEKEYLMVNQKE
15 seq id no 58; SQTSLLEAKKNGLEKEYLMVNQKE
seq id no 59; SQTSLLEAKMA
seq id no 60; SQTSLLEAKMA
seq id no 61; TLVFPK
seq id no 62; KTLVFPK
20 seq id no 63; LKNVEDQKTLVFPK
seq id no 64; LKNVEDQKKTTLVFPK
seq id no 65; LKNVEDQKKH
seq id no 66; LKNVEDQKH
seq id no 67; KKIQLY
25 seq id no 68; KKKIQLY
seq id no 69; RKRF SYTEYLASIIRFIFSVNRRKEIQNLSSCNFKI
seq id no 70; LRIVSYSKKKKIQLY
seq id no 71; LRIVSYSKKKKKIQLY
seq id no 72; LRIVSYSKRKRF SYTEYLASIIRFIFSVNRRKEIQNLS-
30 -SCNFKI
seq id no 73; LRIVSYSKRKRF SYTEYLASIIRFIFSVNRRKEIQNLS-
-SCNFKI
seq id no 74; QDLPLSSICQTIVTIYWQ
seq id no 75; KQDLPLSSICQTIVTIYWQ
35 seq id no 76; NRTCPFRLFVRRMLQFTGNKVLD RP
seq id no 77; GFVVS VVKQDLPLSSICQTIVTIYWQ

- seq id no 78; GFVVSVVKKKQDLPLSSICQTIIVTIYWQ
seq id no 79; GFVVSVVKKNRTPFRLFVRRMLQFTGNKVLDRLP
seq id no 80; GFVVSVVKNRTPFRLFVRRMLQFTGNKVLDRLP
seq id no 81; YRKTKNQ
5 seq id no 82; KYRKTKNQ
seq id no 83; NTERPKIRTN
seq id no 84; DETFYKGKKYRKTKNQ
seq id no 85; DETFYKGKKYRKTKNQ
seq id no 86; DETFYKGKKNTERPKIRTN
10 seq id no 87; DETFYKGKKNTERPKIRTN
seq id no 88; LSINNYRFQMKFYFRFTSHGSPFTSANF
seq id no 89; KLSINNYRFQMKFYFRFTSHGSPFTSANF
seq id no 90; NSVSTTTGFR
seq id no 91; NIQLAATKKLSINNYRFQMKFYFRFTSHGSPFTSANF
15 seq id no 92; NIQLAATKKLSINNYRFQMKFYFRFTSHGSPFTSANF
seq id no 93; NIQLAATKKNSVSTTTGFR
seq id no 94; NIQLAATKNSVSTTTGFR
seq id no 95; MEHVAPGRMSASPQSPTQ
seq id no 96; KMEHVAPGRMSASPQSPTQ
20 seq id no 97; KWSTWLQAECQHLHSPQRSDKPQQAGLDQQHHC FALDS-
-SPGPRPVFLQLLGLMGQGRHD
seq id no 98; WSTWLQAECQHLHSPQRSDKPQQAGLDQQHHC FALDS-
-SPGPRPVFLQLLGLMGQGRHD
seq id no 99; TFSVWAEKMEHVAPGRMSASPQSPTQ
25 seq id no 100; TFSVWAEKKMEHVAPGRMSASPQSPTQ
seq id no 101; TFSVWAEKKWSTWLQAECQHLHSPQRSDKPQQAGLDQ-
-QHHC FALDSSPGPRPVFLQLLGLMGQGRHD
seq id no 102; TFSVWAEKWSTWLQAECQHLHSPQRSDKPQQAGLDQ-
-QHHC FALDSSPGPRPVFLQLLGLMGQGRHD
30 seq id no 103; HKWLKFCLLRLVKESFHE
seq id no 104; KHKWLKFCLLRLVKESFHE
seq id no 105; KGGKAKGKKHKWLKFCLLRLVKESFHE
seq id no 106; KGGKAKGKKHKWLKFCLLRLVKESFHE
seq id no 107; KGGKAKGKNTNG
35 seq id no 108; KGGKAKGKNTNG
seq id no 109; VNNFFKKL

- seq id no 110; KVNFFFKL
- seq id no 111; LSQGNVKKVNFFFKL
- seq id no 112; LSQGNVKKVNFFFKL
- seq id no 113; GEKNDLQLFVMSDRRYKIYWTVILLNPCGNLHLKTTSL
- 5 seq id no 114; KGEKNDLQLFVMSDRRYKIYWTVILLNPCGNLHLKTTSL
- seq id no 115; KGKMICSYS
- seq id no 116; GKMICSYS
- seq id no 117; SSKTFEKKGEKNDLQLFVMSDRRYKIYWTVILLNPCGN-
-LHLKTTSL
- 10 seq id no 118; SSKTFEKKKGEKNDLQLFVMSDRRYKIYWTVILLNPCGN-
-LHLKTTSL
- seq id no 119; SSKTFEKKKGKMICSYS
- seq id no 120; SSKTFEKKKGKMICSYS
- seq id no 121; QRKPKRANCVIQRRAM
- 15 seq id no 122; KQRKPKRANCVIQRRAM
- seq id no 123; NKENQKEQTALLYRGGQRCRCVCLRF
- seq id no 123; NKENQKEQTALLYRGGQRCRCVCLRF
- seq id no 124; PDYQPPAKKQRKPKRANCVIQRRAM
- seq id no 125; PDYQPPAKKKQRKPKRANCVIQRRAM
- 20 seq id no 126; PDYQPPAKKNKENQKEQTALLYRGGQRCRCVCLRF
- seq id no 127; PDYQPPAKKNKENQKEQTALLYRGGQRCRCVCLRF
- seq id no 128; NLSSLLI
- seq id no 129; TCLPF
- seq id no 130; QPTFTLRKNLSSLLI
- 25 seq id no 131; QPTFTLRKNLSSLLI
- seq id no 132; QPTFTLRKKTCLPF
- seq id no 133; QPTFTLRKTCLPF
- seq id no 134; RATFLLSLWECSLPQARLCLIVSRTGLLVQS
- seq id no 135; GQHFYWHCGSAACHRRGCV
- 30 seq id no 136; KENVRDKKRATFLLSLWECSLPQARLCLIVSRTGLLVQS
- seq id no 137; KENVRDKKKRATFLLSLWECSLPQARLCLIVSRTGLLVQS
- seq id no 138; KENVRDKKGQHFYWHCGSAACHRRGCV
- seq id no 139; KENVRDKKGQHFYWHCGSAACHRRGCV
- seq id no 140; ITHTRWGITTWDSWSVRMKANWIAQQNKSLILSPSFTK
- 35 seq id no 141; KITHTRWGITTWDSWSVRMKANWIAQQNKSLILSPSFTK
- seq id no 142; KLLTPGGELPHGILGQ

seq id no 143; LLTPGGELPHGILGQ

seq id no 144; PPVCELEKITHTRWGITTWDSWSVRMKANWIAQQNKS-
-LILSPSFTK

5 seq id no 145; PPVCELEKKITHTRWGITTWDSWSVRMKANWIAQQNKS-
-LILSPSFTK

seq id no 146; PPVCELEKKLLTPGGELPHGILGQ

seq id no 147; PPVCELEKKLLTPGGELPHGILGQ

seq id no 148; SLKDELEKMKI

seq id no 149; SLKDELEKMKI

10 seq id no 150; LGQSSPEKKNKN

seq id no 151; LGQSSPEKKNKN

seq id no 152; RLRRINGRGSQIRSRNAFNRSEE

seq id no 153; EPKVKEEKKT

seq id no 154; EPKVKEEKKKT

15 seq id no 155; EPKVKEEKKRLRRINGRGSQIRSRNAFNRSEE

seq id no 156; EPKVKEEKKRLRRINGRGSQIRSRNAFNRSEE

seq id no 157; TFRYKGKQHPFFST

seq id no 158; GPNAPEEKNH

seq id no 159; GPNAPEEKNH

20 seq id no 160; GPNAPEEKKTFRYKGKQHPFFST

seq id no 161; GPNAPEEKTFRYKGKQHPFFST

seq id no 162; MQNTCV

seq id no 163; KMQNTCV

seq id no 164; KCKIRVFSK

25 seq id no 165; CKIRVFSK

seq id no 166; FFKRTVQKMQNTCV

seq id no 167; FFKRTVQKMQNTCV

seq id no 168; FFKRTVQKCKIRVFSK

seq id no 169; FFKRTVQKCKIRVFSK

30 seq id no 170; LPHYLAH

seq id no 171; CLITWLTN

seq id no 172; GSTTGLSATPLPHYLAH

seq id no 173; GSTTGLSATPPLPHYLAH

seq id no 174; GSTTGLSATPPCLITWLTN

35 seq id no 175; GSTTGLSATPCLITWLTN

seq id no 176; RFADKPRPN

- seq id no 177; DLPTSPDQTRSGPVHVSVEP
seq id no 178; DSAAGCSGTPRFADKPRPN
seq id no 179; DSAAGCSGTPPRFADKPRPN
seq id no 180; DSAAGCSGTPPDLPSTSPDQTRSGPVHVSVEP
5 seq id no 181; DSAAGCSGTPDLPTSPDQTRSGPVHVSVEP
seq id no 182; AHPETPAQNRLRIPCSRREVRSRACKPPGAQGSDER-
-RGKASPGRD CDVRTGRP
seq id no 183; PAHPETPAQNRLRIPCSRREVRSRACKPPGAQGSDER-
-RGKASPGRD CDVRTGRP
10 seq id no 184; RPTRRHPRRIASGSPAVGGR
seq id no 185; VAIRGHPRPPAHPETPAQNRLRIPCSRREVRSRACKP-
-PGAQGSDERRGKASPGRD CDVRTGRP
seq id no 186; VAIRGHPRPPP AHPETPAQNRLRIPCSRREVRSRACKP-
-PGAQGSDERRGKASPGRD CDVRTGRP
15 seq id no 187; VAIRGHPRPPRPTRRHPRRIASGSPAVGGR
seq id no 188; VAIRGHPRPPRPTRRHPRRIASGSPAVGGR
seq id no 189; RGR TSGRSLSCCRRPRCRPAVASRSTAPSPRAGSR-
-RCCLRTSCGAARPRRTR SACGDWVASPPTRSS-
-SRTACGAASPPARSWSAP
20 seq id no 190; GGGHLEEV
seq id no 191; YFGGPDSTPRGRTSGRSLSCCRRPRCRPAVASR-
-STAPSPRAGSRRCCLRTSCGAARPRRTR SACGD-
-WVASPPTRSSSRTACGAASPPARSWSAP
seq id no 192; YFGGPDSTPPRGRTSGRSLSCCRRPRCRPAVASR-
25 -STAPSPRAGSRRCCLRTSCGAARPRRTR SACGDW-
-VASPPTRSSSRTACGAASPPARSWSAP
seq id no 193; YFGGPDSTPPGGGHLEEV
seq id no 194; YFGGPDSTPPGGGHLEEV
seq id no 195; HRVADP
30 seq id no 196; LSQSSELDPPSSR
seq id no 197; LSQSSELDPPPSSR
seq id no 198; LSQSSELDPPHRVADP
seq id no 199; LSQSSELDPPHRVADP
seq id no 200; VILLPEDTPPS
35 seq id no 201; VILLPEDTPPS
seq id no 202; VILLPEDTPPLLRA

- seq id no 203; VILLPELDPLLRA
seq id no 204; PSPLP
seq id no 205; PLLFHRPCSPSPALGATVLAVYRYE
seq id no 206; LLFHRPCSPSPALGATVLAVYRYE
5 seq id no 207; APRPPLGPPSPLP
seq id no 208; APRPPLGPPPSPLP
seq id no 209; APRPPLGPPPLL FHRPCSPSPALGATVLAVYRYE
seq id no 210; APRPPLGPPPLL FHRPCSPSPALGATVLAVYRYE
seq id no 211; TQVLPQGCSLSLLHTTTFPHRQVPHILDW
10 seq id no 212; PTQVLPQGCSLSLLHTTTFPHRQVPHILDW
seq id no 213; PLQSF PKDAASAFSTPRFPTDKFPTSWTGSCPGQPHGT-
-RAFCQPGPEFNAFSAC
seq id no 214; LQSF PKDAASAFSTPRFPTDKFPTSWTGSCPGQPHGT-
-RAFCQPGPEFNAFSAC
15 seq id no 215; PSRPQSQPPTQVLPQGCSLSLLHTTTFPHRQVPHILDW
seq id no 216; PSRPQSQPPTQVLPQGCSLSLLHTTTFPHRQVPHILDW
seq id no 217; PSRPQSQPPPLQSF PKDAASAFSTPRFPTDKFPTS-
-WTGSCPGQPHGTRAFCQPGPEFNAFSAC
seq id no 218; PSRPQSQPPLQSF PKDAASAFSTPRFPTDKFPTS-
20 -WTGSCPGQPHGTRAFCQPGPEFNAFSAC
seq id no 219; TAWPGRRRFTTPEPYCLCTPLGPWAPRFLW
seq id no 220; PTAWPGRRRFTTPEPYCLCTPLGPWAPRFLW
seq id no 221; PRGPAGGALLPRSLTAFVPHSGHGLPVSSGEPAYTPIP-
-HDVPHGTTPPFC
25 seq id no 222; RGPAGGALLPRSLTAFVPHSGHGLPVSSGEPAYTPIPH-
-DVPHTGTPPFC
seq id no 223; DLPAVPGPPTAWPGRRRFTTPEPYCLCTPLGPWAPRFLW
seq id no 224; DLPAVPGPPPTAWPGRRRFTTPEPYCLCTPLGPWAPRFLW
seq id no 225; DLPAVPGPPPRGPAGGALLPRSLTAFVPHSGHGLPVSSG-
30 -EPAYTPIPHDVPHGTTPPFC
seq id no 226; DLPAVPGPPRGPAGGALLPRSLTAFVPHSGHGLPVSSG-
-EPAYTPIPHDVPHGTTPPFC
seq id no 227; QWGLSWMS
seq id no 228; NGDCHGCPEGRQSL
35 seq id no 229; FTMDRVLTPQWGLSWMS

seq id no 230; FTMDRVLTPPQWGLSWMS
seq id no 231; FTMDRVLTPPNGDCHGCPEGRQSL
seq id no 232; FTMDRVLTPPNGDCHGCPEGRQSL
seq id no 233; HHPARQCPHCIMHLQTQLIHRNLTGPSQLTSLHRS-
5 -PYQIAATPWTTDFAASFFLNPVTPFLLCRRCQGKDV-
-LCTNARCLSQTSPSHHKALSRTTTQCMNT-
-TPWLAVRPAKAFPLL
seq id no 234; PHHPARQCPHCIMHLQTQLIHRNLTGPSQLTSLHRS-
-PYQIAATPWTTDFAASFFLNPVTPFLLCRRCQGK-
10 -DVLCTNARCLSQTSPSHHKALSRTTTQCMNTTP-
-WLAVRPAKAFPLL
seq id no 235; HTIQHASVPTASCISKLNSYTEN
seq id no 236; PQVGMRPSNP PHHPARQCPHCIMHLQTQLIHRNLT-
-GPSQLTSLHRSPYQIAATPWTTDFAASFFLNPVTPFL-
15 -LCRRCQGKDVLCNARCLSQTSPSHHKALSRTTTQC-
-MNTTPWLAVRPAKAFPLL
seq id no 237; PQVGMRPSNP PHHPARQCPHCIMHLQTQLIHRNLTGPS-
-QLTSLHRSPYQIAATPWTTDFAASFFLNPVTPFLLCRRC-
-QGKDVLCNARCLSQTSPSHHKALSRTTTQCMNTTPWLA-
20 -VRPAKAFPLL
seq id no 238; PQVGMRPSNP HTIQHASVPTASCISKLNSYTEN
seq id no 239; PQVGMRPSNP HTIQHASVPTASCISKLNSYTEN
seq id no 240; WAARSWCERRAAVAPLAPWAWGCPAGCTPPVAARAC-
-AATRPEGWRSPCTH
25 seq id no 241; PWAARSWCERRAAVAPLAPWAWGCPAGCTPPVAA-
-RACAATRPEGWRSPCTH
seq id no 242; RGLRGAGARGGLRLLRHLRPGLGDALRGVHPPLR-
-LGPALLPAPRGGEAPAHTDARARRVHGAGGDRGHGPAAL
seq id no 243; EEKLARCRPPWAARSWCERRAAVAPLAPWAWGCPAGC-
30 -TPPVAARACAATRPEGWRSPCTH
seq id no 244; EEKLARCRPPWAARSWCERRAAVAPLAPWAWGCPA-
-GCTPPVAARACAATRPEGWRSPCTH
seq id no 245; EEKLARCRPPRGLRGAGARGGLRLLRHLRPGLGDA-
-LRGVHPPLRLGPALLPAPRGGEAPAHTDARARRVHGAGG-
35 -DRGHGPAAL

- seq id no 246; EEKLARCRPRGLRGAGARGGLRLLRHLRPGLGDALRG-
-VHPPLRLGPALLPAPRGGEAPAHTDARARRVHGAGG-
-DRGHPGPAAL
- 5 seq id no 247; QPPVSPRRRPGRPRAPPPPQPMVSPRRRTTGPPW-
-RPPPLQSTMSPPPQALHQAQLLWCTTAPLPGLPQPQ-
-PARALHSQFPATTLILLPPLPAIAPRLMPVALTIARYL-
-LSPPPITALLPSCLLGSLSFSCFLTFTQSSLIPLW-
-KIPAPTTTKSCRETFWK
- 10 seq id no 248; SPGCHLGPGDQAAPGLHRPPSPWCHLGAGQQARLGVHR-
-PSSPQCHLGLRLCIRLSFYSGAQRHLCQGYHNPSQQEHS-
-ILNSQPPL
- 15 seq id no 249; KPAPGSTAPQPPVSPRRRPGRPRAPPPPQPMVSPRR-
-RTTGPPWRPPPLQSTMSPPPQALHQAQLLWCTTAP-
-LPGLPQPQPARALHSQFPATTLILLPPLPAIAPRLMPVA-
-LTIARYLLSPPPITALLPSCLLGSLSFSCFLTFTQTS-
-SLIPLWKIPAPTTTKSCRETFWK
- 20 seq id no 250; KPAPGSTAPPQPPVSPRRRPGRPRAPPPPQPMVSPR-
-RRTTGPPWRPPPLQSTMSPPPQALHQAQLLWCT-
-TAPLPGLPQPQPARALHSQFPATTLILLPPLPAIAP-
-RLMPVALTIARYLLSPPPITALLPSCLLGSLSFSCFLT-
-TFQTSSLIPLWKIPAPTTTKSCRETFWK
- 25 seq id no 251; KPAPGSTAPSPGCHLGPGDQAAPGLHRPPSPWCHL-
-GAGQQARLGVHRPSSPQCHLGLRLCIRLSFYSGA-
-QRHLCQGYHNPSQQEHSILNSQPPL
- 30 seq id no 252; KPAPGSTAPSPGCHLGPGDQAAPGLHRPPSPWCHL-
-GAGQQARLGVHRPSSPQCHLGLRLCIRLSFYSGAQ-
-RHLCQGYHNPSQQEHSILNSQPPL
- 35 seq id no 253; QPMVSPRRRTTGPPWRPPPLQSTMSPPPQALHQAQ-
-LLWCTTAPLPGLPQPQPARALHSQFPATTLILLPPLP-
-AIAPRLMPVALTIARYLLSPPPITALLPSCLLGSL-
-SFSCFLTFTQSSLIPLWKIPAPTTTKSCRETFWK
- seq id no 254; SPWCHLGAGQQARLGVHRPSSPQCHLGLRLCIRLSF-
-YSGAQRHLCQGYHNPSQQEHSILNSQPPL
- seq id no 255; RPPPGSTAPQPMVSPRRR
- seq id no 256; RPPPGSTAPPQPMVSPRRR
- seq id no 257; RPPPGSTAPPSPWCHLGA

seq id no 258; RPPPGSTAPSPWCHLGA
seq id no 259; RPRAPPPSPWCHL
seq id no 260; RPRAPPPSPWC
seq id no 261; RPRAPPPAHGVTSAP
5 seq id no 262; RPRAPPPAHGV
seq id no 263; APGLHRPPQPMVSP
seq id no 264; AAPGLHRPPQPMVSPR
seq id no 265; PGLHRPPAHGV
seq id no 266; APGLHRPPAHGVTS
10 seq id no 267; VDRPQHTEWLSWSNLYRIRHQ
seq id no 268; HYLCTDVAPR
seq id no 269; HYLCTDVAPPR
seq id no 270; HYLCTDVAPPVDRPQHTEWLSWSNLYRIRHQ
seq id no 271; HYLCTDVAPVDRPQHTEWLSWSNLYRIRHQ
15 seq id no 272; SAYLSPLGTTWLRTCACRLPRPAASCLCTTPSLLW-
-PRRTCAGSPRATSSPWRMPAPKSCCTTGLAFTS-
-PIGLGWSATASGYARIWPVLSLTCQSWSTSLPSTAVTW
seq id no 273; PSAYLSPLGTTWLRTCACRLPRPAASCLCTTPSLLWP-
-RRTCAGSPRATSSPWRMPAPKSCCTTGLAFTSP-
20 -IGLGWSATASGYARIWPVLSLTCQSWSTSLPSTAVTW
seq id no 274; PAPIFLLWGPLG
seq id no 275; APIFLLWGPLG
seq id no 276; LPARAPGPPSAYLSPLGTTWLRTCACRLPRPAASCL-
-CTTPSLLWPRRTCAGSPRATSSPWRMPAPKSCC-
25 -TTGLAFTSPIGLGWSATASGYARIWPVLSLT-
-QSWSTSLPSTAVTW
seq id no 277; LPARAPGPPPSAYLSPLGTTWLRTCACRLPRPAAS-
-CLCTTPSLLWPRRTCAGSPRATSSPWRMPAPKSCC-
-TTGLAFTSPIGLGWSATASGYARIWPVLSLTC-
30 -QSWSTSLPSTAVTW
seq id no 278; LPARAPGPPAPIFLLWGPLG
seq id no 279; LPARAPGPPAPIFLLWGPLG
seq id no 280; DLEHHGGVTRHRHR
seq id no 281; LVSDYSMTPRP
35 seq id no 282; LVSDYSMTPPRP
seq id no 283; LVSDYSMTPPDLEHHGGVTRHRHR

seq id no 284; LVSDYSMTPDLEHHGGVTRHRHR

seq id no 285; FHHIATDVGPVVRIGFLKIKGKIKGKSLRKPNW-
-KTQHKLKRALMFLIVKKL

5 seq id no 286; PFHHIATDVGPVVRIGFLKIKGKIKGKSLRKPNWK-
-TQHKLKRALMFLIVKKL

seq id no 287; PSITLQOMLAPS

seq id no 298; SITLQOMLAPS

10 seq id no 289; TSCNEMNPPFHHIATDVGPVVRIGFLKIKGKIKGSL-
-RKPNWKTQHKLKRALMFLIVKKL

seq id no 290; TSCNEMNPPFHHIATDVGPVVRIGFLKIKGKIKG-
-KSLRKPNWKTQHKLKRALMFLIVKKL

seq id no 291; TSCNEMNPPSITLQOMLAPS

seq id no 292; TSCNEMNPPSITLQOMLAPS

15 seq id no 293; LEMILFLMTF

seq id no 294; HPCITKTFLEMILFLMTF

seq id no 295; HPCITKTFFLEMILFLMTF

seq id no 296; HPCITKTFFWR

seq id no 297; HPCITKTFFWR

20 seq id no 298; LMFESQMRLNSKNAHLPIISF

seq id no 299; EYGSIIAFLMFESQMRLNSKNAHLPIISF

seq id no 300; EYGSIIAFLMFESQMRLNSKNAHLPIISF

seq id no 301; HLNKGRRLGDKIRAT

seq id no 302; FHLNKGRRLGDKIRAT

25 seq id no 303; VTSGTPFFHLNKGRRLGDKIRAT

seq id no 304; VTSGTPFFFHLNKGRRLGDKIRAT

seq id no 305; VTSGTPFFFI

seq id no 306; VTSGTPFFFI

seq id no 307; CEIERIHFFF

30 seq id no 308; CEIERIHFFSK

seq id no 309; CEIERIHFSK

seq id no 310; FRYISKSI

seq id no 311; RYISKSI

seq id no 312; FKKYEPIFFRYISKSI

35 seq id no 313; FKKYEPIFRYISKSI

- seq id no 314; FPDSQPGPLYPLDPSCLISSASNPQELSDCHYIH-
-LAFGFSNWRSCPVLPGHCGVQ
- seq id no 315; PDSQPGPLYPLDPSCLISSASNPQELSDCHYIHL-
-AFGFSNWRSCPVLPGHCGVQ
- 5 seq id no 316; LNMFAVFS
seq id no 317; LNMFAVFFS
seq id no 318; LNMFAVFFPDSQPGPLYPLDPSCLISSASNPQE-
-LSDCHYIHLAFGFSNWRSCPVLPGHCGVQ
- 10 seq id no 319; LNMFAVFFPDSQPGPLYPLDPSCLISSASNPQELS-
-DCHYIHLAFGFSNWRSCPVLPGHCGVQ
seq id no 320; AMEETVVAVATVETEVEAMEETGVVAAMEETEVGAT-
-EETEVEAMEAKWEEETTTEMISATDHT
seq id no 321; LWVRPWLWEWLRWRPKWRLWRRQEWRLWRRPRWGL-
-RRRPRWLWRENGRKKRLQK
- 15 seq id no 322; YGGDRSRGAMEETVVAVATVETEVEAMEETGVVAAM-
-EETEVGATEETEVEAMEAKWEEETTTEMISATDHT
seq id no 323; YGGDRSRGGAMEETVVAVATVETEVEAMEETGVVA-
-AMEETEVGATEETEVEAMEAKWEEETTTEMISATDHT
seq id no 324; YGGDRSRGGLWVRPWLWEWLRWEPKWRLWRRQEW-
20 -RLWRRPRWGLRRRPRWLWRENGRKKRLQK
seq id no 325; YGGDRSRGLWVRPWLWEWLRWEPKWRLWRRQEW-
-LWRRPRWGLRRRPRWLWRENGRKKRLQK
seq id no 326; EFGGRRQK
seq id no 327; EFGGRRQK
- 25 seq id no 328; RRAKGGGAGASNPRQ
seq id no 329; GRRAKGGGAGASNPRQ
seq id no 330; DVGLREGALELPTRGNKRNV
seq id no 331; MRGGGGVGGRRAKGGGAGASNPRQ
seq id no 332; MRGGGGVGGRRAKGGGAGASNPRQ
- 30 seq id no 333; MRGGGGVGGDVGLREGALELPTRGNKRNV
seq id no 334; MRGGGGVGGDVGLREGALELPTRGNKRNV
seq id no 335; VWQLAGPMLAGWRSLSWFCRMYGI
seq id no 336; CGSWPALCWRAGGVWAVGSAGCMEYDPEALPAWGP-
-AAAATVHPRR
- 35 seq id no 337; RRYPCWGVWQLAGPMLAGWRSLSWFCRMYGI

- seq id no 338; RRYPCWGGVWQLAGPMLAGWRSLSWFCRMYGI
seq id no 339; RRYPCWGGCGSWPALCWRAGGVWAVGSAGCMEYD-
-EALPAAWGPAAAATVHPRR
seq id no 340; RRYPCWGGCGSWPALCWRAGGVWAVGSAGCMEYDPE-
5 -ALPAAWGPAAAATVHPRR
seq id no 341; LWLWAGWTVWWSCGPGEKGHWPSLPTMALLLRFSM-
-RVASY
seq id no 342; GLWLWAGWTVWWSCGPGEKGHWPSLPTMALLL-
-RFSCMRVASY
10 seq id no 343; GCGCPAGQYGGAVGLARRGTAGCLPCPPWLCCCCAF-
-PACGLPGTDGWRGWQSGCVRVSGSAPWAPGFPFSP-
-PCPLCGTQPRW
seq id no 344; CGCGPAGQYGGAVGLARRGTAGCLPCPPWLCCCCAF-PACG-
-LPGTDGWRGWQSGCVRVSGSAPWAPGFPFSP-
15 -PLCGTQPRW
seq id no 345; LAFNVPGGGLWLWAGWTVWWSCGPGEKGHWPSLPTMA-
-LLLLRFSCMRVASY
seq id no 346; LAFNVPGGGLWLWAGWTVWWSCGPGEKGHWPSLPTM-
-ALLLRFSMVRVASY
20 seq id no 347; LAFNVPGGCGCGPAGQYGGAVGLARRGTAGCLPCPP-
-WLCCCCAF-PACGLPGTDGWRGWQSGCVRVSGSAPW-
-APGFPFSP-PCPLCGTQPRW
seq id no 348; LAFNVPGGCGCGPAGQYGGAVGLARRGTAGCLPCPPW-
-LCCCCAF-PACGLPGTDGWRGWQSGCVRVSGSAPWA-
25 -PGFPFSP-PCPLCGTQPRW
seq id no 349; PPMMPGQREAPGRQEA
seq id no 350; GPPMPGQREAPGRQEA
seq id no 351; GHQCQCQKGKGRHRADRRPDTAQEE
seq id no 352; HQCQCQKGKGRHRADRRPDTAQEE
30 seq id no 353; GGHSYGGGPPMPMPGQREAPGRQEA
seq id no 354; GGHSYGGGPPMPMPGQREAPGRQEA
seq id no 355; GGHSYGGGHQCQCQKGKGRHRADRRPDTAQEE
seq id no 356; GGHSYGGGHQCQCQKGKGRHRADRRPDTAQEE
seq id no 357; APCPQSSGGG
35 seq id no 358; LPAPSQAAADELDRRPG

- seq id no 359; TKVRLIRGAPCPQSSGGG
seq id no 360; TKVRLIRGGAPCPQSSGGG
seq id no 361; TKVRLIRGGLPAPSQAAADELDRRPG
seq id no 362; TKVRLIRGLPAPSQAAADELDRRPG
5 seq id no 363; CSLAKDGSTEDTVSSLCGEEDTEDEELEAAASHLNK-
-DLYRELLGG
seq id no 364; GCSLAKDGSTEDTVSSLCGEEDTEDEELEAAASHLNK-
-DLYRELLGG
seq id no 365; AAAWQKMAPPRTPRPACVARR
10 seq id no 366; ENSRPKRGGCSLAKDGSTEDTVSSLCGEEDTEDEELE-
-AAASHLNKDLYRELLGG
seq id no 367; ENSRPKRGGCSLAKDGSTEDTVSSLCGEEDTEDE-
-ELEAAASHLNKDLYRELLGG
seq id no 368; ENSRPKRGGAAAWQKMAPPRTPRPACVARR
15 seq id no 369; ENSRPKRGAAAWQKMAPPRTPRPACVARR
seq id no 370; HCVLAASGAS
seq id no 371; GHCVLAASGAS
seq id no 372; GTASSRPLGLPKPHLHRPVPIRHPSCP
seq id no 373; TASSRPLGLPKPHLHRPVPIRHPSCP
20 seq id no 374; AGTLQLGGHCVLAASGAS
seq id no 375; AGTLQLGGGHCVLAASGAS
seq id no 376; AGTLQLGGGTASSRPLGLPKPHLHRPVPIRHPSCP
seq id no 377; AGTLQLGGTASSRPLGLPKPHLHRPVPIRHPSCP
seq id no 378; RRTPSTEKR
25 seq id no 379; RRTPSTEKKR
seq id no 380; RRTPSTEKKGRSEC
seq id no 381; RRTPSTEKGRSEC
seq id no 382; STTKCQSGTAETYNSWKVKNLQLEPRRVTSQMNQVK-
-DMTAILSQS
30 seq id no 384; SSEEIKKKSTTKCQSGTAETYNSWKVKNLQLEPRRV-
-TSQMNQVKDMTAILSQS
seq id no 385; SSEEIKKKSTTKCQSGTAETYNSWKVKNLQLEPRR-
-VTSQMNQVKDMTAILSQS
seq id no 386; SSEEIKKKVQPNASQAQQKPTTHGR
35 seq id no 387; SSEEIKKKVQPNASQAQQKPTTHGR

seq id no 388; NRGWVGAGE
seq id no 389; IEAG
seq id no 390; VHNYCNMKNRGWVGAGE
seq id no 391; VHNYCNMKKNRGWVGAGE
5 seq id no 392; VHNYCNMKKIEAG
seq id no 393; VHNYCNMKIEAG
seq id no 394; QLRCWNTWAKMFFMVFLIIWQNTMF
seq id no 395; VKKDNHKKQLRCWNTWAKMFFMVFLIIWQNTMF
seq id no 396; VKKDNHKKQLRCWNTWAKMFFMVFLIIWQNTMF
10 seq id no 397; VKKDNHKKKNS
seq id no 398; VKKDNHKKNS
seq id no 399; GAEESGPFNRQVQLKVHASGMGRHLWNCPAFWSEV
seq id no 400; HPSPPPEKRS
seq id no 401; HPSPPPEKKRS
15 seq id no 402; HPSPPPEKKGAEESGPFNRQVQLKVHASGMGRHLW-
-NCPAFWSEV
seq id no 403; HPSPPPEKGAEEGPFNRQVQLKVHASGMGRHLWN-
-CPAFWSEV
seq id no 404; MQVLSKTHMNLFPQVLLQMFLRGLKRLQLDLEKSKKRKL
20 seq id no 405; RCKSARLI
seq id no 406; VQTQPAIKKMQVLSKTHMNLFPQVLLQMFLRGLKRLQL-
-DLEKSKKRKL
seq id no 407; VQTQPAIKKMQVLSKTHMNLFPQVLLQMFLRGLKRL-
-LQDLEKSKKRKL
25 seq id no 408; VQTQPAIKKRCKSARLI
seq id no 409; VQTQPAIKRCKSARLI
seq id no 410; ARSGKKQKRKL
seq id no 411; ARSGKKQKKRKL
seq id no 412; ARSGKKQKKENFS
30 seq id no 413; ARSGKKQKENFS
seq id no 414; KASARSGKSKKRKL
seq id no 415; KASARSGKSKKRKL
seq id no 416; KASARSGKAKKENSF
seq id no 417; KASARSGKAKKENSF
35 seq id no 418; HLNKGRRLGDKIRAT
seq id no 419; VTSGTPFFHLNKGRRLGDKIRAT

- seq id no 420; VTSGTPFFFHLNKGRRRLGDKIRAT
 seq id no 421; VTSGTPFFFI
 seq id no 422; VTSGTPFFI
 5 seq id no 423; VTLLYVNTVTLAPNVNMESSRNAHSPATPSAKRK-
 -DPDLTWGGFVFFFCQFH
 seq id no 424; KCRCKPNFFVTLLYVNTVTLAPNVNMESSRNAHSP-
 -ATPSAKRKDPDLTWGGFVFFFCQFH
 seq id no 425; KCRCKPNFFVTLLYVNTVTLAPNVNMESSRNAH-
 -SPATPSAKRKDPDLTWGGFVFFFCQFH
 10 seq id no 426; KCRCKPNFFL
 seq id no 427; KCRCKPNFL
 seq id no 429; LVKKLKEKKMNWIL
 seq id no 430; LVKKLKEKKMNWIL
 seq id no 431; LVKKLKEKKR
 15 seq id no 432; LVKKLKEKR
 seq id no 433; AAIVKDCCR
 seq id no 434; SQPASILGRKL
 seq id no 435; SQPASILGKRKL
 seq id no 436; SQPASILGAAIVKDCCR
 20 seq id no 437; SQPASILGAAIVKDCCR
 seq id no 459; NTWAKMFFMVFLIIWQNTMF

25 Examples of cancers particularly suitable for treatment
 with one or a combination of several of this compounds
 are: colorectal cancer, breast cancer, small-cell lung
 cancer, non small-cell lung cancer, liver cancer (primary
 and secondary), renal cancer, melanoma, ovarian cancer,
 30 cancer of the brain, head and neck cancer, pancreatic
 cancer, gastric cancer, eosophageal cancer, prostate
 cancer and leukemias and lymphomas.

Below are listed some examples of where these mutations
 may result in gene products that result in development of
 35 tumours:

Development of colorectal cancers are believed to result from a series of genetic alterations. Deleted in colorectal cancer (DCC) gene (seq id nos 30-34), Human cysteine protease (ICErel-III) gene (seq id nos 394-398 and 459), Human putative mismatch repair /binding protein (hMSH3) gene (Seq id hos 134-147), Human hMSH6 gene (seq id nos 201-204 and 295-299), Human n-myc gene (seq id nos 190-195), Human TGF β 2 (hTGF β 2) gene (seq id nos 22-29), Human p53 associated gene (seq id nos 287-294) may be involved in colorectal cancer.

Human breast cancer susceptibility (BRCA2) (seq id nos 35-94) and Human BRCA1-associated RING domain protein (BARD1) gene (seq id nos 404-413) are involved in breast cancer and ovarian cancer Human hMSH6 gene (seq id nos 201-204 and 295-299) may be involved in brain tumours.

Gene alteration are frequent in many types of adenocarcinomas , below are listed some genes that are mutated in many cancers:

Human breast cancer susceptibility (BRCA2) gene (seq id nos 35-94), Deleted in colorectal cancer (DCC) gene (seq id nos 30-34), Human putatative mismatch repair/binding protein (hMSH3) gene (seq id nos 134-147), Human hMSH6 gene (seq id nos 201-204 and 295-299), human N-MYC gene (seq id no 190-195), Human TGF β 2 (hTGF β 2) gene (seq id nos 22-29), Human p53 associated gene (seq id nos 287-294), Human MUC1 gene (seq id nos 248-267), Human germline n-myc gene (seq id nos 184-195), Human Wilm's tumour (WIT-1) associated protein (seq id nos 388-393), Human nasopharynx carcinoma EBV BNLF-1 gene (seq id nos 205-211), Human transforming growth factor-beta induced gene product (BIGH3) seq id nos 228-233).

Many of the mutated genes may result in development of leukemias and lymphomas: Human neurofibromin (NF1) gene (seq id nos 178-183), b-raf oncogene (seq id nos 172-177), Human protein-tyrosine kinase (JAK1) gene (seq id nos 268-272), Human protein-tyrosine kinase (JAK3) gene (seq id nos 273-280) are examples.

Genes involved in malignant melanoma: Human malignant melanoma metastasis-supressor (hKiSS-1) gene (seq id nos 331-337), Genes involved in metastasis: Human metastasis-associated mta1 (hMTA1) gene (seq id nos 360-365).

Cell cycle control and signal transduction is strickly regulated. Frameshift mutations in these genes may result in uncontrolled cell growth. Examples of genes which may be suseptable are: Human protein tyrosine phosphatase (hPTP) gene (seq id nos 95-102), Human kinase (TTK) gene (seq id nos 109-121), Human transcriptional repressor (CTCF) gene (seq id nos 122-128), Human cell cycle regulatory protein (E1A-binding protein) p300 gene (seq id nos 212-219), Human tranforming growth factor-beta induced gene product (BIGH3) (seq id nos 228-233), Human FLt4 gene (for transmembrane tyrosinase kinase (seq id nos 281-286), Human G protein-coupled receptor (hGPR1) gene (seq id nos 317-322), Human transcription factor (hITF-2) gene (seq id nos 329-330), Human telomerase-associated protein TP-1 (hTP-1) gene (seq id nos 338-351), Human transcription factor TFIIB 90 kDa subunit (hTFBIIB90) gene (seq id nos 366-373), Human FADD-homologous ICE/CED-3like protease gene (seq id nos 129-133)

Mutations in DNA synthesis or -repair enzymes may also lead to uncontrolled cell growth. Human DNA topoisomerase II (top2) gene (seq id nos 103-108) and Human putative mismatch repair/binding protein (hMSH3) gene (seq id nos

134-147) and (hMSH6) gene (seq id nos 201-204 and 205-299).

5 The following are tumour suppressor genes, Human retinoblastoma binding protein 1 isoform I (hRBP1) gene (seq id nos 148-158), Human neurofibromin(NF1) gene (seq id nos 178-183), Human p53 associated gene (seq id nos 287-294), Human retinoblastoma related protein (p107) gene (seq id nos 312-316), Human tumour suppressor (hLUCA-1) gene (seq id nos 374-381), Mutations in these genes may
10 result in development of cancer.

The following are oncogenes, proto-oncogenes or putative oncogenes ; Human germline n-myc gene (seq id nos 184-189), Human n-myc gene (seq id nos 190-195), Human can (hCAN) gene (seq id nos 300-302), Human dek (hDEK) gene (seq id nos 309-311), b-raf oncogene (seq id nos 172-177), Human DBL (hDBL) proto-oncogene / Human MCF2PO (hMCF2PO) gene (seq id nos 303-308). Frameshift mutations in these
15 genes may lead to development of cancer.
20

BIOLOGICAL EXPERIMENTS

25 Description of the Figures

FIG. 1:

It has been demonstrated that T cells from normal donors can be stimulated with a mixture of peptides containing
30 both mutant BAX and mutant TGF β RII peptides. Peptide mixture dependent T cell proliferation in blood samples from six different donors are shown in figure 1. The results were obtained by stimulating peripheral blood mononuclear cells (PBMCs) from each donor with a mixture of mutant BAX peptides (seq id nos 1,9-12) and mutant TGF β RII peptides (seq id nos 15-21). The concentration of each

individual peptide in the mixture was 20 μ M. After two weeks, and weekly thereafter, the bulk cultures were restimulated with autologous PBMCs pulsed with 10-25 μ M of the peptide mixture. After 4-5 restimulations the bulk cultures were tested in a standard proliferation assay with PBMCs alone or as a control or PBMCs pulsed with 25 μ M of the peptides as antigen presenting cells (APCs).

FIG. 2:

It has further been found that T cell clones can be generated against separate peptides of the mixture used in the bulk stimulation experiments. Figure 2 shows the proliferation of T cell clone 521-2 which was obtained by cloning the bulk culture from donor 1 (figure 1) by seeding 5 cells per well in U-bottomed, 96-well microtiter plates and using autologous PBMCs pulsed with 25 μ M of the mutant BAX peptide with seq id no 12 as feeder cells. Autologous B-lymphoblastoid cells were used as APCs in the proliferation assay.

FIG. 3:

In figure three it is shown that mutant BAX peptides and mutant TGF β R11 peptides can be used to stimulate T cells (PBMCs) from a patient with breast cancer. Dendritic cells (DCs) from the same cancer patient were used as APCs. The T cell stimulation (figure 3) was obtained by pulsing DCs separately with a mixture of mutant BAX peptides (seq id nos 1,9-12) and a mixture of mutant TGF β R11 peptides (seq id nos 15-21) followed by addition of autologous PBMCs and 10 ng/ml tumour necrosis factor. The concentration of each peptide in the mixtures used for pulsing was 25 μ M. The PBMCs and the DCs were obtained by leukapheresis from a patient with breast cancer who had been on a granulocyte colony stimulating factor (G-CSF) treatment. The CD34+

cells were isolated from the cell product before DCs were derived using standard methods.

FIG. 4:

5 Figure 4 shows the capability of T cells obtained from ascites fluid of a pancreatic cancer patient to recognise and proliferate to different synthetic peptides derived from mutant BAX (seq id nos 1,9-12) and mutant TGF β R11 (seq
10 id nos 15,17-21). The T cell line was obtained after expansion of T cells present in the ascites fluid of a patient with pancreatic adenocarcinoma. The T cell line was expanded in vitro by culturing with 100 U/ml recombinant interleukin-2 (rIL-2) (Amersham, Aylesbury, UK) for one
15 week before being tested in a proliferation assay.

Autologous, irradiated (30Gy) PBMCs were seeded 5×10^4 in u-bottomed 96-well plates (Costar, Cambridge, MA) and pulsed with single synthetic peptides at 20 μ M for 2h. The
20 T cells were added 5×10^4 per well and the plates were incubated for four days at 37°C with addition of 18.5×10^4 Bq/mL 3H-thymidine for the last 12 hours before harvesting. The plates were counted in a liquid scintillation counter (Packard Topcount). Data represent specific proliferation
25 to the different synthetic peptides and values are expressed as the mean of triplicate cultures. These results show that T cells isolated from a pancreatic cancer patient are capable of responding to a panel of peptides carrying amino acid sequences derived from mutant BAX and TGF β R11.

30 FIG. 5:

Figure 5 further demonstrates the capability T cells from another pancreatic cancer patient to recognise and
35 proliferate to different synthetic peptides derived from mutant BAX and mutant TGF β R11. The T cell line was obtained after expansion of T cells present in the ascites fluid of

a patient with pancreatic adenocarcinoma. The experiment was set up in the same way as described above. Data represent specific proliferation to the different synthetic peptides and values are expressed as the mean of triplicate
5 cultures.

In order to investigate the T cell response from the latter pancreatic cancer patient, responding T cells were cloned. Peritoneal macrophages were irradiated (30 Gy) and plated 1
10 $\times 10^4$ into U-bottomed 96-well plates (Costar) together with 25 μM of each peptide. T cell blasts were counted in a microscope and added 5 blasts per well together with 100 U/ml human recombinant interleukin-2 (rIL-2) (Amersham, Aylesbury, UK) in a total volume of 200 μL . After 14 days T
15 cell clones were transferred onto 24-well plates (Costar) with 1 mg/mL phytohemagglutinin (PHA, Wellcome, Dartford, UK), 100 U/ml rIL-2 and allogeneic, irradiated PBMCs as feeder cells and screened for peptide specificity after 7 and 14 days.

20

FIG. 6:

T cell clone 520.5, 520.7 and 520.8 were selected for further characterisation and express the cell surface phenotype CD3+, CD8+ and TcR+. Figure 6 shows the
25 recognition and cytotoxicity of T cell clone 520.5, 520.7 and 520.8 against peptide-pulsed autologous target cells pulsed with the seq id no 10 peptide. Autologous Epstein-barr virus transformed B-cells (EBV) were labelled with 3H-thymidine (9.25×10^4 Bq/ml) over night, washed
30 once and plated 2500 cells per well in 96-well plates with or without 25 μM of synthetic peptide (seq id no 10) and 1% DMSO in medium. After 30 minutes incubation at 37°C the plates were washed before addition of T cells. The plates were further incubated at 37°C for 4 hours and then
35 harvested before counting in a liquid scintillation counter (Packard Topcount). Data represent percent specific lysis

of 3H-thymidine labelled peptide pulsed target cells at an effector/target ratio of 10/1. Values are expressed as the mean of triplicate cultures. These results demonstrate that the three different T cell clones obtained from ascites fluid of a pancreatic carcinoma patient, exhibit specific cytotoxicity of autologous EBV targets pulsed with the relevant peptide (seq id no 10) derived from mutant BAX.

FIG. 7:

Figure 7 shows the cytolytic properties of three different T cell clones obtained from the same patient. These T cell clones were cultured and expanded as described above, but they were generated against a synthetic peptide the seq id no 17 peptide carrying amino acid sequences derived from mutant TGF β R11. T cell clone 538.1, 538.3 and 538.4 all show the cell-surface phenotype CD3+, CD8+ and TcR+. The experimental conditions were as described above (figure 6). Data represent percent specific lysis of 3H-thymidine labelled peptide pulsed target cells pulsed with the seq id no 428 peptide at an effector/target ratio of 10/1. Values are expressed as the mean of triplicate cultures. These results demonstrate that the three different T cell clones obtained from ascites fluid of a pancreatic carcinoma patient, exhibit specific cytotoxicity of autologous EBV targets pulsed with the relevant peptide (seq id no 428) derived from mutant TGF β R11.

FIG. 8:

Figure 8 shows the specificity of two CD4+ T cell clones, IMT8 and IMT9, obtained from a tumour biopsy taken from a patient with an adenocarcinoma localised to the proximal colon. Immunohistochemistry revealed that the patient had an abundant infiltrate of predominantly CD4+ T cells, many of which carried activation markers. In areas of CD4 T cell infiltration islands of HLA DR positive tumour cells were observed. The T cell clones were obtained from the

component of tumour infiltrating lymphocytes which grew out of the biopsy following culture in medium containing 15 U/ml of recombinant human IL-2 for 16 days. The T cells from this culture were cloned by limiting dilution (1 cells/well) in Terasaki plates with irradiated peptide pulsed APC and 100 U/ml of IL-2. Pulsing of autologous APC was performed with a mixture of the TGF β RII frameshift peptides with sequence identity no. 15, 17 and 18 at 1 μ g/ml of each peptide in the presence of 3 μ g/ml of purified human β 2 microglobulin and 10ng/ml of recombinant human TNF α for 3 hrs at 37 °C. Of the 14 clones that could be expanded preliminary tests showed that two of the clones were reactive with the peptide mixture used for cloning. After expansion the clones were screened for reactivity with the single peptides in a standard proliferative assay. The results show that IMT8 and IMT9 both react specifically with the TGF β RII frameshift peptide with seq. id. no. 17, no reactivity was observed with the two other frameshift peptides tested.

The figure (Fig. 8) depicts the results of conventional T cell proliferative assays, where cloned T cells (5×10^4) and irradiated APC (5×10^4) were cocultured for 3 days in triplicates before harvesting. To measure the proliferative capacity of the cultures, ^3H -thymidine ($3,7 \times 10^4$ Bq/well) was added to the culture overnight before harvesting) Values are given as mean counts per minute (cpm) of the triplicates.

FIG. 9:

Figure 9 demonstrates that the specific reactivity of the two T cell clones IMT8 and IMT9 against the peptide with seq. id.no. 17 is completely blocked by treatment of the cells with an antibody that specifically binds to HLA-DR molecules, since the reactivity after blocking is the same

as the background reactivity of the clones with APC in the absence of the peptide. On the other hand antibodies to the HLA class II isotypes HLA-DQ and -DP failed to block the reactivity of the clones with peptide pulsed APC. This experiment unequivocally identifies HLA-DR as the molecule responsible to present the peptide to these two T cell clones. Antibody blocking experiments were performed using the homozygous EBV transformed cell line 9061 (IHWS9 nomenclature) as APC. The APC were pulsed with peptide at a concentration of 15 µg/ml for 1 hr at 37 °C before addition of blocking antibodies L243 (pan-DR antibody), SPVL3 (pan-DQ antibody) and B7.21 (pan-DP antibody) at 10 µg/ml. Unpulsed APC and APC pulsed with peptide in the absence of blocking antibody served as negative and positive controls respectively. Results are expressed as in Figure 8.

FIG. 10:

The patient IMT was HLA typed and turned out to be HLA: A1,2; B7,8; DR3,14; DQ1,2. To determine which of the HLA-DR molecules that were responsible for presentation of the peptide with seq. id. no. 17, a panel of HLA workshop derived homozygous BCL cell lines were obtained and pulsed with the peptide with seq. id. no. 17. Figure 10 describes the identification of HLA-DR14 (DRA*0102, DRB*1401) as the HLA-DR molecule responsible for presentation of the peptide with seq. id. no. 17 to the T cell clones IMT8 and IMT9. A specific proliferative response was observed when peptide was presented by the autologous EBV transformed cell line (Auto APC) and by cell lines 9054 (EK) and 9061 (31227ABO), both of which expressed DR14 as the only DR molecule on their surface. The homozygous cell line gave higher responses, reflecting a higher level of expression of the relevant class II/peptide complexes due to the effect of a double dose of the genes encoding this DR molecule. No response was obtained when the peptide was presented by cell lines expressing HLA-DR3 (9018, LOO81785), which

represents the other DR molecule expressed by the patients APC, nor by irrelevant HLA-DR molecules. The experiment was performed as described in figure 9, with the exception that no antibody blocking was performed. Results are expressed
5 as in Figure 8.

FIG. 11:

Figure 11 describes the dose response curves obtained by pulsing the cell line 9054 with increasing concentrations
10 of the peptide with seq. id. no. 17. Both IMT 8 and IMT9 demonstrate a dose dependent increase in the proliferative response to the peptide. Results were performed as described in Fig. 9 and 10 with the peptide concentrations indicated on the Figure (Fig. 11). Results are expressed as
15 in Fig.8.

FIG. 12:

Figure 12 describes the reactivity of a cell line generated by in vitro stimulation of T cells isolated from peripheral
20 blood from a healthy blood donor (Donor 2892) by weekly stimulation with irradiated autologous dendritic cells pulsed with the peptides with sequence identity numbers 16, 17 and 21. A specific response above background values was obtained when the T cells were co-incubated with autologous
25 dendritic cells pulsed with the peptide with seq. id. no. 21. No activity could be detected in the culture after the first and second in vitro stimulation. These data demonstrate that the T cell repertoire of normal individuals contain a few precursor cells that have the
30 capacity to recognise this frameshift peptide derived from a mutation in TGF β RII that does not occur in normal people. In two other blood donors (#2706 and #2896), the level of precursor cells with the relevant specificity was too low to be detected. The results are expressed as spots per 10⁴
35 T cells tested in a conventional IFNg ELISPOT assay. This assay enumerates the number of cells present in a mixture

of cells that are capable of specifically reacting with a defined antigen. Briefly 10^7 T cells (non adherent cells) were stimulated weekly with $2-5 \times 10^6$ irradiated peptide pulsed autologous dendritic cells (DC) as APC. The DC were generated from the adherent cell population by culture for one week in recombinant human GM-CSF and IL-4 according to standard protocols as described in the literature. After peptide pulsing overnight at $15 \mu\text{g/ml}$ of peptide, full maturation of the DC was obtained by culture with recombinant TNF α . ELISPOT was performed according to standard published protocols using 10^4 cultured T cells per well in duplicate and 10^4 peptide pulsed or unpulsed DC as APC. The results are expressed as mean number of spots per 10^4 T cells.

FIG. 13:

Figure 13 shows the results of in vitro stimulation of T cells from a healthy blood donor (Donor 322) with peptides with sequence identity number 15-21. In vitro culture was performed as described in Figure 12. A proliferative response above background values was seen when the T cell culture primed with a mixture of the peptides with seq. id. no. 16 and 21 was stimulated with peptide 21 and the culture primed with the peptide with seq. id. no. 17 was stimulated with the same peptide. These results demonstrate that normal blood donors have small numbers of circulating T cells specific for these frameshift peptides, and that it is possible to expand these cells in culture by stimulation with frameshift peptides. These results also confirmed the results shown in Fig. 8-11, demonstrating that the peptide with seq. id. no. 17 is immunogenic in humans, and indicate that the peptide with seq. id. no. 21 may also be used as a cancer vaccine in humans. The results are expressed as described in Fig. 8.

FIG. 14:

The results shown in Figure 14 demonstrate that CD8+ T cells specific for HLA class I epitopes can be generated from T cells present in the T cell repertoire of a healthy blood donor (donor 905). No reactivity above background was seen with any of the peptides after the second round of in vitro restimulation. After the fourth restimulation, the frequency of T cells specific for the peptide with seq. id. no. 428 had increased from undetectable levels to approximately 2,5 % of the cells. These results demonstrate that CTL precursors of the CD8+ phenotype are present in the unprimed T cell repertoire of healthy blood donors. Such T cells may be expanded in vitro by specific stimulation with the peptide with seq. id. no. 428. This forms the basis for using this peptide as a cancer vaccine to elicit cytotoxic T cells specific for frameshift peptides in cancer patient having such mutations. T cells were generated by weekly restimulation of T cells isolated from peripheral blood and stimulated with peptide pulsed autologous DC as described in Fig.12, with the exception that IL-7 and IL-2 was added during culture according to standard procedures for generating cytotoxic T cells (CTL) of the CD8 phenotype. The peptides used were peptides with sequence identity number 428, 439, 446 and 451. Cells were tested in ELISPOT assay as described in Fig.12. The results are expressed as described in figure 12.

The peptide with seq. id. no. 17 was selected and designed to contain binding motifs for both several HLA class I and HLA class II molecules. These peptides thus contain epitopes both for CD4+ and CD8+ T cells, and was predicted to elicit both CD4 and CD8 T cell responses in cancer patient provided processing of the aberrant TGF β RII protein naturally occurring in cancer cells would take place and

result in an overlapping peptide. This has now been proven for CD4 T cells by the results in Fig. 8-11. These results have the following implication:

- 5 1) The results in Figure 8 prove that the mutated form of TGF β RII Receptor which occurs in a high proportion of cancer patients with defects in their mismatch repair machinery is a tumour specific antigen.
- 10 2) The antigen specificity of the infiltrating T cells commonly observed in colorectal cancer are generally not known. The results in figure 8 demonstrate that one component of the T cells constituting the population of tumour infiltrating lymphocytes in this patients tumour is
15 specific for a frameshift mutation, demonstrating that TGF β RII frameshift peptides are immunogenic *in vivo*, occasionally giving rise to spontaneous T cell activation.
- 20 3) It follows from this observation that processing of the non-functional form of the TGF β RII Receptor that is formed by the common frameshift mutation is processed. This processing may take place either in the tumour cell as part of natural breakdown of the aberrant protein, or after the tumour cell itself or a released form of the receptor has
25 been taken up by a professional APC or both.
- 30 4) The results in Figure 8 also indicate that the peptide with seq. id. no. 17 is capable of binding to an HLA class II molecule, since pulsing of APC with this peptide results in a specific proliferative response against the peptide,
35 and since CD4 T cell responses always are class II restricted. That this is the case is demonstrated by the results of the experiment shown in Figure 9. Here it is shown that the specific response against the peptide with seq. id. no. 17 is completely blocked by an antibody to

HLA-DR, but not with antibodies to the two other HLA class II molecules, HLA-DQ and -DP. Furthermore, by using a panel of standard homozygous Epstein Barr Virus (EBV) transformed B Cell Lines (BCL) covering the relevant HLA class II

5 molecules present on the patients own APC, we were able to identify the class II molecule responsible for presentation of the peptide with seq. id. no. 17 to TLC IMT8 and IMT9 as being HLA-DR 14. Together these findings fit extremely

10 well with the immunohistological observations made in parallel sections taken from the same tumour biopsy, where we could show that activated CD4+ T cells were abundant in the proximity of tumour cells that had been induced to express HLA-DR. molecules. The results in Figure 11 demonstrate that these T cell clones are capable of

15 mounting a proliferative response over a range of peptide doses and that the responses are dose dependent.

5) Since these T cell clones were obtained by cloning T cells isolated from a tumour biopsy, another implication of

20 our finding is that activated T cells specific for the peptide with seq. id. no. 17 are capable of homing to the tumour tissue after activation.

6) Since the peptide with seq. id. no. 17 is a tumour

25 specific antigen, and since frameshift mutations giving rise to this peptide or peptides with overlapping sequences are commonly found in cancers with defects in enzymes that are part of the mismatch repair machinery, this peptide may be used as a vaccine to elicit T cell response in cancer

30 patients or patients at high risk for developing cancer. Such T cell responses may potentially influence the growth of an existing tumour or prohibit regrowth of tumour after surgery and other forms of treatment or be given to patients with an inheritable form of cancer where a defect

35 mismatch enzyme is detected or suspected and that have a

high chance of developing a cancer where this precise mismatch repair mutation will occur.

5

Synthesis

The peptides were synthesised by using continuous flow solid phase peptide synthesis. N-a-Fmoc-amino acids with appropriate side chain protection were used. The Fmoc-amino acids were activated for coupling as pentafluorophenyl esters or by using either TBTU or diisopropyl carbodiimide activation prior to coupling. 20% piperidine in DMF was used for selective removal of Fmoc after each coupling. Cleavage from the resin and final removal of side chain protection was performed by 95% TFA containing appropriate scavengers. The peptides were purified and analysed by reversed phase (C18) HPLC. The identity of the peptides was confirmed by using electro-spray mass spectroscopy (Finnigan mat SSQ710).

The peptides used for *in vitro* studies of T cell stimulation were synthesised by this method.

Several other well known methods can be applied by a person skilled in the art to synthesise the peptides.

Examples of the method for determining new frameshift mutation peptides.

In this Example, the BAX gene is used to illustrate the principle.

In each of the steps listed below, the 1st line is the gene sequence and 2nd line is amino acid sequence.

In the steps 2-5, the outlined sequences represent the mutant part of the protein.

5 Step one:

Normal BAX.

10 ATG GGG GGG GAG GCA CCC GAG CTG GCC CTG GAC CCG GTG
M G G E A P E L A L D P V ...

15 Step two:

1G deleted from gene sequence.

20 ATG GGG GGG AGG CAC CCG AGC TGG CCC TGG ACC CGG TGC CTC
M G G R H P S W P W T R C L

AGG ATG CGT CCA CCA AGA AGC TGA
R M R P P R S stop

25

Step three:

30 2G deleted from gene sequence.

ATG GGG GGA GGC ACC CGA GCT GGC CCT GGA CCC GGT GCC
M G G G T R A G P G P G A

35 TCA GGA TGC GTC CAC CAA GAA GCT GAG CGA GTG TCT CAA GCG
S G C V H Q E A E R V S Q A

40 CAT CGG GGA CGA ACT GGA CAG TAA
H R G R T G Q stop

Step four:

1G inserted in gene sequence.

5 ATG GGG GGG GGA GGC ACC CGA GCT GGC CCT GGA CCC GGT GCC
 M G G G G T R A G P G P G A
 TCA GGA TGC GTC CAC CAA GAA GCT GAG CGA GTG TCT CAA GCG
 10 S G C V H Q E A E R V S Q A
 CAT CGG GGA CGA ACT GGA CAG TAA
 H R G R T G Q stop

15

Step five:

2G inserted in gene sequence.

20 ATG GGG GGG GGG AGG CAC CCG AGC TGG CCC TGG ACC CGG TGC
 M G G G R H P S W P W T R C
 CTC AGG ATG CGT CCA CCA AGA AGC TGA
 25 L R M R P P R S stop

In the next Example, the TGF β RII gene is used to illustrate the principle.

30

In each of the steps listed below, the 1st line is the gene sequence and 2nd line is amino acid sequence.

In the steps 2-5, the outlined sequences represent the mutant part of the protein.

35

Step one:

Normal TGF β RII.

40

GAA AAA AAA AAG CCT GGT GAG ACT TTC TTC ATG TGT TCC....
 E K K K P G E T F F M C S...

45

Step two:

1A deleted from gene sequence.

5 GAA AAA AAA AGC CTG GTG AGA CTT TCT TCA TGT GTT CCT GTA
 E K K S L V R L S S C V P V

10 GCT CTG ATG AGT GCA ATG ACA ACA TCA TCT TCT CAG AAG AAT
 A L M S A M T T S S S Q K N

15 ATA ACA CCA GCA ATC CTG ACT TGT TGC TAG
 I T P A I L T C C stop

Step three:

2A deleted from gene sequence.

20 GAA AAA AAA GCC TGG TGA
 E K K A W stop

Step four:

1A inserted in gene sequence.

30 GAA AAA AAA AAA GCC TGG TGA
 E K K K A W stop

Step five:

2A inserted in gene sequence.

35 GAA AAA AAA AAA AGC CTG GTG AGA CTT TCT TCA TGT GTT CCT
 E K K K S L V R L S S C V P

40 GTA GCT CTG ATG AGT GCA ATG ACA ACA TCA TCT TCT CAG AAG
 V A L M S A M T T S S S Q K

45 AAT ATA ACA CCA GCA ATC CTG ACT TGT TGC TAG
 N I T P A I L T C C stop

Thus the peptides of the invention may be used in a
 50 method for the treatment of cancers with cancer cells
 harbouring genes with frameshift mutations, which

treatment comprises administering at least one peptide of the present invention *in vivo* or *ex vivo* to a human patient in need of such treatment.

- 5 In another embodiment the peptides of the invention may be used to vaccinate a human being disposed for cancers with cancer cells harbouring genes with frameshift mutations, by administering at least one peptide of the present invention to said human being.

10

It is further considered to be an advantage to administer to a human individual a mixture of the peptides of this invention, whereby each of the peptides of the invention can bind to different types of HLA class I and/or class

15

II molecules of the individual.

It is further anticipated that the power of an anticancer vaccine or peptide drug as disclosed in the above mentioned PCT/NO92/00032 application, can be greatly

20

enhanced if the peptides of the present invention were included. Thus in another embodiment of the present invention peptides of the present invention are administered together with, either simultaneously or in optional sequence, with the peptides disclosed in

25

PCT/NO92/00032.

It is considered that the peptides may be administered together, either simultaneously or separately, with compounds such as cytokines and/or growth factors, i.e. interleukin-2 (IL-2), interleukin-12 (IL-12), granulocyte

30

macrophage colony stimulating factor (GM-CSF), Flt-3 ligand or the like in order to strengthen the immune response as known in the art.

35

The peptides according to the present invention can be used in a vaccine or a therapeutical composition either

alone or in combination with other materials, such as for instance standard adjuvants or in the form of a lipopeptide conjugate which as known in the art can induce high-affinity cytotoxic T lymphocytes, (K. Deres, 5 Nature, Vol.342, (nov.1989)).

The peptides according to the present invention may be useful to include in either a peptide or recombinant fragment based vaccine.

10

The peptides according to the present invention can be included in pharmaceutical compositions or in vaccines together with usual additives, diluents, stabilisers or the like as known in the art.

15

According to this invention, a pharmaceutical composition or vaccine may include the peptides alone or in combination with at least one pharmaceutically acceptable carrier or diluent.

20

Further a vaccine or therapeutical composition can comprise a selection of peptides which are fragments of the mutant proteins arising from insertion or deletion of bases in a repeat sequence of the gene.

25

Further a vaccine composition can comprise at least one peptide selected for one cancer, which vaccine would be administered to a person carrying a genetic disposition for this particular cancer.

30

Further a vaccine composition can comprise at least one peptide selected for one cancer, which vaccine would be administered to a person belonging to a high risk group for this particular cancer.

35

The cancer vaccine according to this invention may further be administered to the population in general for example as a mixture of peptides giving rise to T cell immunity against various common cancers connected with frameshift mutation genes.

The peptides according to this invention may be administered as single peptides or as a mixture of peptides. Alternatively the peptides may be covalently linked with each other to form larger polypeptides or even cyclic polypeptides.

A cancer therapy according to the present invention may be administered both in vivo or ex vivo having as the main goal the raising of specific T cell lines or clones against the mutant gene product associated with the cancer type with which the patient is afflicted.

Further, the frameshift mutant peptides of this invention may be administered to a patient by various routes including but not limited to subcutaneous, intramuscular, intradermal, intraperitoneal, intravenous or the like. In one embodiment the peptides of this invention are administered intradermally. The peptides may be administered at single or multiple injection sites to a patient in a therapeutically or prophylactically effective amount.

The peptides of this invention may be administered only once or alternatively several times, for instance once a week over a period of 1-2 months with a repeated sequence later all according to the need of the patient being treated.

The peptides of this invention can be administered in an amount in the range of 1 microgram (1 μ g) to 1 gram (1g)

to an average human patient or individual to be vaccinated. It is preferred to use a smaller dose in the range of 1 microgram (1 µg) to 1 milligram (1 mg) for each administration.

5

The invention further encompasses DNA sequences which encodes a frameshift mutation peptide.

10

The invention additionally encompasses isolated DNA sequences comprising a DNA sequence encoding at least one frameshift mutant peptide, and administration of such isolated DNA sequences as a vaccine for treatment or prophylaxis of cancers associated with frameshift mutations in the genes.

15

The peptides according to this invention may be administered to an individual in the form of DNA vaccines. The DNA encoding these peptides may be in the form of cloned plasmid DNA or synthetic oligonucleotide. The DNA may be delivered together with cytokines, such as IL-2, and/or other co-stimulatory molecules. The cytokines and/or co-stimulatory molecules may themselves be delivered in the form of plasmid or oligonucleotide DNA. The response to a DNA vaccine has been shown to be increased by the presence of immunostimulatory DNA sequences (ISS). These can take the form of hexameric motifs containing methylated CpG, according to the formula:

20

5'-purine-purine-CG-pyrimidine-pyrimidine-3'. Our DNA vaccines may therefore incorporate these or other ISS, in the DNA encoding the peptides, in the DNA encoding the cytokine or other co-stimulatory molecules, or in both. A review of the advantages of DNA vaccination is provided by Tighe et al (1998, *Immunology Today*, 19(2), 89-97).

30

35 In one embodiment, the DNA sequence encoding the mutant BAX peptides comprises:

Normal BAX.

5 ATG GGG GGG GAG GCA CCC GAG CTG GCC CTG GAC CCG GTG

10 1G deleted from BAX gene sequence.

ATG GGG GGG AGG CAC CCG AGC TGG CCC TGG ACC CGG TGC CTC
AGG ATG CGT CCA CCA AGA AGC TGA

15

2G deleted from BAX gene sequence.

20 ATG GGG GGA GGC ACC CGA GCT GGC CCT GGA CCC GGT GCC
TCA GGA TGC GTC CAC CAA GAA GCT GAG CGA GTG TCT CAA GCG
CAT CGG GGA CGA ACT GGA CAG TAA

25

1G inserted in BAX gene sequence.

30 ATG GGG GGG GGA GGC ACC CGA GCT GGC CCT GGA CCC GGT GCC
TCA GGA TGC GTC CAC CAA GAA GCT GAG CGA GTG TCT CAA GCG
CAT CGG GGA CGA ACT GGA CAG TAA

35

2G inserted in BAX gene sequence.

40 ATG GGG GGG GGG AGG CAC CCG AGC TGG CCC TGG ACC CGG TGC
CTC AGG ATG CGT CCA CCA AGA AGC TGA

45

In a second embodiment, the DNA sequence encoding the
mutant TGF β R11 peptides comprises:

50

Normal TGF β RII gene.

GAA AAA AAA AAG CCT GGT GAG ACT TTC TTC ATG TGT TCC....

5

1A deleted from TGF β RII gene sequence.

GAA AAA AAA AGC CTG GTG AGA CTT TCT TCA TGT GTT CCT GTA
10 GCT CTG ATG AGT GCA ATG ACA ACA TCA TCT TCT CAG AAG AAT
ATA ACA CCA GCA ATC CTG ACT TGT TGC TAG

15

2A deleted from TGF β RII gene sequence.

GAA AAA AAA GCC TGG TGA

20

1A inserted in TGF β RII gene sequence.

25

GAA AAA AAA AAA GCC TGG TGA

30

2A inserted in TGF β RII gene sequence.

GAA AAA AAA AAA AGC CTG GTG AGA CTT TCT TCA TGT GTT CCT
35 GTA GCT CTG ATG AGT GCA ATG ACA ACA TCA TCT TCT CAG AAG
AAT ATA ACA CCA GCA ATC CTG ACT TGT TGC TAG

40

The invention further encompasses vectors and plasmids comprising a DNA sequence encoding a frameshift mutant peptide. The vectors include, but are not limited to E.Coli plasmid, a Listeria vector and recombinant viral
45 vectors. Recombinant viral vectors include, but are not limited to orthopox virus, canary virus, capripox virus, suipox virus, vaccinia, baculovirus, human adenovirus,

SV40, bovine papilloma virus and the like comprising the DNA sequence encoding a frameshift mutant peptide.

5 It is considered that an anticancer treatment or prophylaxis may be achieved also through the administration of an effective amount of a recombinant virus vector or plasmid comprising at least one insertion site containing a DNA sequence encoding a frameshift mutant peptide to a patient, whereby the patient's
10 antigen presenting cells are turned into host cells for the vector/plasmid and presentation of HLA/frameshift mutation peptide complex is achieved.

15 A person skilled in the art will find other possible use combinations with the peptides of this invention, and these are meant to be encompassed by the present claim.

The peptides according to this invention may be produced by conventional processes as known in the art, such as
20 chemical peptide synthesis, recombinant DNA technology or protease cleavage of a protein or peptide encoded by a frameshift mutated gene. One method for chemical synthesis is elucidated in the description below.

25 In order for a cancer vaccine and methods for specific cancer therapy based on specific T cell immunity to be effective, three conditions must be met:
1. The peptides used must correspond, either in their full length or after processing by antigen presenting
30 cells, to the processed mutant protein fragment as presented by a HLA Class I and/or class II molecule on the cancer cell or other antigen presenting cells,
2. The peptides used must be bound to a HLA Class I and/or Class II molecule in an immunogenic form, and

3. T-cells capable of recognising and responding to the HLA/peptide complex must be present in the circulation of the human being.

5 It has been established that all these conditions are met for some representative peptides according to the present invention. The peptides according to the present invention give rise to specific T cell immune responses in vitro. It has been established that the peptides according to this invention correspond to processed mutant protein fragments. This is exemplified with peptides corresponding to fragments of transformed mutant BAX and TGF β RII peptides.

15 Through the present invention the following advantages are achieved:

- It offers a possibility to treat patients suffering from cancers arising from frame-shift mutations in their genes, most of which cancers known at present do not have any good treatment alternatives.

20 - It offers a possibility to vaccinate prophylactically humans carrying genetic dispositions or belonging to other high risk groups.

- It offers a possibility to prepare a combination treatment for a specific cancer, such as for instance colorectal or pancreatic cancers, wherein the cancer commonly is associated with either a frameshift mutation or a point mutation in the genes.

30 -Since described frameshift mutations occurs in a large variety of cancers it will be possible to use this peptides in combination with established vaccines and future vaccines to obtain a multiple targetting treatment.

-Likewise patients suffering from cancers associated with multiple frameshift mutations in genes can be treated more efficiently through a combination treatment.